REVIEW



WT1 gene: a potential therapeutic target for multiple cancer treatment strategies

Tetsuyuki Hirahata¹ · Reeshan Ul Quraish¹ · Afraz Ul Quraish¹ · Shahan Ul Quraish¹ · Reezan Ul Quraish¹

Received: 5 March 2025 / Accepted: 6 June 2025 © The Author(s) 2025

Abstract

The Wilms' Tumor 1 (WT1) gene has attracted significant attention in oncological research owing to its involvement in the pathogenesis of a variety of malignancies. Initially identified in association with Wilms' tumor, a pediatric renal carcinoma, WT1 is now recognized for its broader implications in the etiology of multiple cancers, including leukemia, ovarian cancer, and lung cancer. This review offers a comprehensive analysis of the historical development of WT1 research, explores its mechanisms in cancer progression, and assesses its potential as a therapeutic target, with an emphasis on its clinical implications for cancer treatment strategies.

Keywords WT1 gene · Tumor suppressor gene · Oncogene · Gene therapy · Immunotherapy

Introduction

Cancer remains one of the most significant global health challenges, contributing to high rates of morbidity and mortality. It ranks among the top three leading causes of death in individuals aged 30 to 69 years in 177 out of 183 countries [1]. According to the 2022 Global Cancer Statistics, nearly 20 million new cancer cases and approximately 10 million cancer-related deaths occurred worldwide in that year [2]. Cancer encompasses a diverse group of diseases characterized by uncontrolled and abnormal cell proliferation. These malignant cells can invade surrounding tissues and metastasize to distant organs via the blood and lymphatic systems. Metastasis is the predominant cause of cancer-related deaths [3–5]. Normal cellular development and tissue homeostasis rely on tightly regulated processes, including cell proliferation, differentiation, and apoptosis. Disruption in any of these processes can result in unchecked cell growth, contributing to cancer initiation and progression [6]. Cell growth is regulated through the coordinated actions of specific genes within each cell. Genomic instability plays a central role in

Two major categories of genes are implicated in cancer: tumor suppressor genes and oncogenes. A disruption or imbalance between these gene classes can drive abnormal cell proliferation. Oncogenes (OGCs) arise from mutations in proto-oncogenes—genes that normally regulate cell division and growth. When mutated, proto-oncogenes become oncogenes, leading to excessive protein production or increased activity beyond normal regulatory limits. This dysregulation enables cells to proliferate autonomously, disregarding the organism's growth control mechanisms.

Tumor suppressor genes (TSGs) are essential for maintaining normal cellular homeostasis by regulating cell division and promoting apoptosis. Under physiological conditions, TSGs prevent excessive cell proliferation and ensure the removal of damaged or abnormal cells. However, when TSGs become inactivated or dysfunctional, cells can proliferate uncontrollably, bypassing the body's regulatory mechanisms. The simultaneous activation of oncogenes (OGCs) and inactivation of TSGs create a permissive environment for unchecked cell growth and resistance to apoptosis—two fundamental hallmarks of cancer initiation and progression [8–10].

Advancing the identification, functional characterization, and mechanistic understanding of these gene classes is critical for progress in cancer research and clinical management. This knowledge underpins the discovery of

Published online: 31 October 2025



the development of neoplastic diseases by promoting mutations and chromosomal alterations [7].

Reeshan Ul Quraish reeshan.17@yahoo.com; reeshan@hicc.jp

Hirahata Gene Therapy Laboratory, HIC Clinic #1105. Itocia Office Tower 11F, 2-7-1. Yurakucho, Chiyoda-Ku, Tokyo 100-0006, Japan

novel biomarkers and drives the development of targeted therapies, ultimately enabling improved prevention, early detection, and personalized treatment strategies.

The WT1 gene (Wilms Tumor 1) was first identified in studies of Wilms tumor, a pediatric renal malignancy. In 1990, researchers mapped the WT1 gene to chromosome 11p13 by analyzing chromosomal abnormalities in affected patients. These findings led to the recognition of WT1 as a tumor suppressor gene, based on its frequent inactivation in Wilms tumor cases [11].

Wilms' tumor is the most common pediatric renal malignancy, accounting for approximately 7% of all child-hood cancers [12]. Genetic analyses of affected patients frequently reveal mutations in several genes, including the homozygous inactivation of the WT1 tumor suppressor gene [13, 14].

Despite being initially identified and classified as a tumor suppressor, accumulating research has demonstrated that WT1 can also function as an oncogene under certain cellular contexts, earning it the designation of a "chameleon gene." [15]. The significance of WT1 extends beyond oncology; it plays critical roles in developmental biology, cancer progression, and clinical diagnostics. Recent studies, such as that by Jing et al. (2022), have shown that WT1 can inhibit the proliferation of human renal carcinoma cells and induce G2/M cell cycle arrest by upregulating IL-24 expression [16].

Moreover, WT1 is implicated in the pathogenesis of a broad spectrum of malignancies, including hematologic cancers and various solid tumors. These include leukemia, breast cancer, ovarian cancer, renal cell carcinoma, mesothelioma, melanoma, glioblastoma, and soft tissue sarcomas, among others [17–24].

Structure of WT1 gene

The human WT1 gene, located on chromosome 11p13, is a complex locus spanning approximately 50-kb and composed of 10 exons. It encodes a 3-kb mRNA transcript [25, 26].

The WT1 gene encodes a transcription factor characterized by four zinc finger motifs at its C-terminal region, each comprising two cysteine and two histidine residues (Fig. 1). These zinc finger domains enable DNA and RNA binding, direct nuclear localization, mediate protein-protein interactions and regulate gene transcription. In contrast, the N-terminal region is enriched in proline, glutamic acid, serine, and glycine residues. This region enables WT1 to self-associate and plays a crucial role in regulating both transcriptional activation and repression [27–29]. The WT1 gene generates multiple transcript variants through alternative splicing at two coding exons. One splicing event involves the inclusion or exclusion of exon 5, which encodes a 17-amino acid peptide, located just N-terminal to the four zinc finger motifs. Another splicing event occurs at exon 9, resulting in the inclusion or exclusion of a three amino acid sequence lysine-threonine-serine (KTS)-positioned between the third and fourth zinc fingers [30–33]. Alternative splicing produces several WT1 isoforms, with four major variants designated A, B, C, and D. Isoform A lacks both the 17-amino acid insert and the KTS sequence. Isoform B contains the 17-amino acid insert but lacks the KTS sequence.

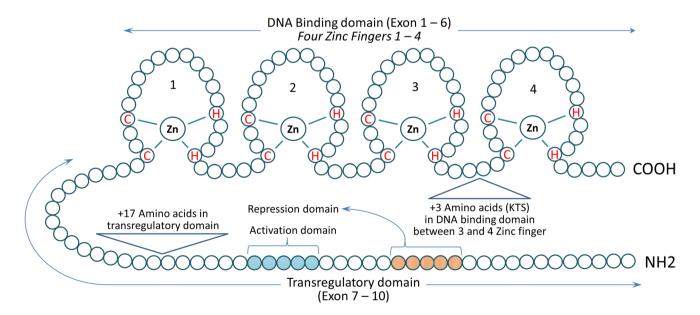


Fig. 1 Schematic Representation of the Structural Organization of the WT1 Protein. The C-terminus of the WT1 protein contains four zinc finger motifs, which are implicated in DNA and RNA binding. The

N-terminus features a transregulatory domain. Brown circles represent the repression domain, while blue circles indicate the activation domain



Isoform C includes the KTS sequence but does not have the 17–amino acid insertion, while isoform D harbors both the 17–amino acid insert and the KTS inserts. These isoforms exhibit distinct functional properties and regulate specific target genes, with expression patterns that are consistent yet tissue-dependent [32, 34, 35].

Molecular function of WT1 gene

WT1 regulates a broad spectrum of target genes involved in key cellular pathways such as growth signaling, cell cycle progression, differentiation, and apoptosis. By modulating these genes, WT1 plays critical roles in physiological processes including cell proliferation, survival, programmed cell death, and the maintenance of mesenchymal-epithelial balance. Its transcriptional activity is context-dependent; in some cases, WT1 functions as an activator, while in others, it acts as a repressor. This dual role highlights WT1's versatility in regulating gene expression and cellular processes [36–42]. For instance, WT1 activates Wnt4 gene expression in the kidney to promote MET, while repressing its expression in the heart to regulate EMT. This context-dependent regulation is achieved through WT1's recruitment of different cofactors: coactivators like CBP/p300 in the kidney and corepressors like BASP1 in the heart. [43]. Multiple studies have shown that WT1's dual role as both an activator and repressor is influenced by the diversity of its isoforms, its interactions with different binding partners, and the specific cellular context in which it functions [27, 28, 44]. This functional versatility arises from the WT1 N-terminal region, which contains both a transcriptional activation domain and a suppression domain (SD), allowing it to modulate gene expression in a context-dependent manner. Importantly, a defined region within the N-terminus can inhibit WT1's own activation domain, further contributing to its regulatory complexity [45, 46]. The suppression domain of WT1 represses the activity of other transcriptional activators by recruiting the cosuppressor, brain acid-soluble protein 1 (BASP1). BASP1 serves as a crucial component of the WT1 cosuppressor complex and plays a key role in modulating WT1's transcriptional activity [47, 48].

The WT1 gene and its isoforms regulate genes critical for cellular differentiation and survival in multiple organs, including the spleen, adrenal glands, liver, diaphragm, gonads, kidneys, and urogenital system [49–51]. WT1 is also essential for heart development and regulates gene expression in cardiomyocytes to maintain cardiac homeostasis and promote repair after injury, highlighting WT1 as a promising target for cardiac regeneration therapies [52]. Quenneville et al. (2024) identified a novel WT1 isoform induced under prolonged severe hypoxia in cancer cells, which drives a non-canonical epithelial—mesenchymal transition, illustrating WT1's functional versatility through isoform-specific

gene regulation in stress conditions [53]. Additionally, studies have demonstrated that loss of WT1 function impairs the regulation of key genes required for kidney development and promotes tumorigenesis, contributing to the onset of Wilms' tumor. These observations emphasize WT1's critical role in regulating cellular proliferation and differentiation [54].

WT1 also contributes significantly to RNA processing and metabolism by interacting with splicing factors. In addition, it influences mRNA transport and stability. [32, 44, 55]. WT1 participates in translation process, as the WT1 protein shuttles between the nucleus and cytoplasm [56, 57]. Moreover, it regulates mRNA turnover through interactions with the 3' untranslated region (UTR), implicating microRNA processing pathway genes in the pathogenesis of Wilms' tumor [58].

WT1 exhibits a dual role in cancer pathogenesis

In Wilms' tumor, WT1 primarily functions as a tumor suppressor by regulating genes essential for cellular differentiation and apoptosis. It suppresses oncogenic signaling pathways while promoting differentiation and programmed cell death in tumor cells [59-61]. However, the role of WT1 in other cancers is far more complex and context-dependent, influenced by the cellular environment and its interaction with key pathways such as p53. For instance, Yao et al. (2021) demonstrated that WT1 suppresses acute myeloid leukemia cell proliferation in a p53-dependent manner, highlighting its tumor-suppressive function when p53 is active [62]. Additionally, Jing et al. (2022) reported that WT1 inhibits the proliferation of renal carcinoma cells and induces G2/M arrest by upregulating pro-apoptotic genes IL-24, reinforcing its tumor-suppressive activity in renal cell carcinoma (RCC) [16].

WT1 exhibits a dual role in tumorigenesis, acting as both a tumor suppressor and an oncogene. This functional dichotomy is primarily driven by specific WT1 isoforms, which exert distinct and often opposing effects on key biological processes. Isoform expression levels and activity critically influence tumor progression and clinical outcomes, emphasizing the need for isoform-specific regulation in cancer biology [59, 63]. WT1's oncogenic or tumor-suppressive function also depends on factors such as developmental stage, cell type, differentiation status, and the timing of its expression or mutation. Additionally, its activity is modulated by binding partners, co-occurring genetic mutations, and features of the tumor microenvironment [15, 25, 27, 28, 44].

WT1 predominantly acts as an oncogene in many adult cancers by modulating apoptotic pathway in a complex, context-specific manner. It promotes cell survival by upregulating anti-apoptotic genes and suppressing pro-apoptotic pathways. This oncogenic activity is particularly evident in



leukemic cells, where WT1 fosters an anti-apoptotic environment by enhancing the expression of survival genes and repressing apoptosis, thereby contributing to its oncogenic role [64–67]. Furthermore, mutations in the WT1 gene or epigenetic modifications, such as promoter methylation, can alter its transcriptional activity and shifting its role from tumor suppressive to oncogenic. The epigenetic landscape of the tumor microenvironment further modulates WT1 expression and activity, reinforcing its dual and context-dependent role in cancer pathogenesis [59, 63].

Table 1 presents the context-dependent cellular and tissue-specific functions of the WT1 gene. Originally identified as a tumor suppressor, WT1 also functions as an oncogene in various malignancies. In cancers such as acute myeloid leukemia (AML), myelodysplastic syndromes (MDS), nonsmall-cell lung cancer (NSCLC), and glioblastoma, WT1 promotes cell proliferation and survival by upregulating oncogenic targets, including BCL-2, MYC, VEGF, and EGFR [65, 66, 68, 69]. In these contexts, WT1 serves as a therapeutic target, particularly through immunotherapeutic strategies such as WT1 peptide vaccines. Conversely, in cancers like Wilms' tumor, mesothelioma, and breast cancer, WT1 acts as a tumor suppressor. Loss-of-function mutations in WT1 in these cases are associated with impaired differentiation, reduced apoptosis, and deregulated cell cycle control. Under normal conditions, WT1 supports differentiation, apoptosis, and cell cycle arrest by activating targets like p21, p53, and IGF-binding proteins [15, 70]. These dual and often contradictory roles of WT1 in cancer underscore the importance of context-specific analysis in both diagnostic assessment and therapeutic development.

WT1 interactions with other tumor suppressors

WT1 engages in complex and context-dependent interactions with other tumor suppressor genes, thereby regulating critical cellular processes such as apoptosis, cell cycle progression, and genomic stability. Recent studies demonstrate that WT1 directly interacts with members of the p53 protein

family including p53, p63, and p73—modulating their transcriptional activities and influencing cellular outcomes related to apoptosis, cell cycle control, and tumorigenesis (Tables 2 and 3).

P53:

WT1 physically interacts with p53 through its zinc finger domains, particularly zinc fingers 1 and 2 [71]. This interaction stabilizes p53 by extending its half-life and shielding it from proteasomal degradation. As a consequence, WT1 enhances p53's transcriptional activity, promoting the expression of genes involved in cell cycle arrest and apoptosis [42, 44]. Despite this activation, WT1 paradoxically inhibits p53-induced apoptosis, highlighting a contextdependent regulatory complexity in their interaction [44, 71]. Bordin et al. (2018) further confirmed this intricate relationship by showing that WT1 loss impairs p53-mediated DNA damage responses in T-cell acute lymphoblastic leukemia, reducing transcription of pro-apoptotic genes such as BAX, BBC3, and GADD45A [72]. Additionally, another study provided evidence that p53 can transcriptionally repress WT1 target genes such as podocalyxin, revealing reciprocal regulation between these tumor suppressors [73]. Collectively, these studies illustrate that the WT1-p53 interaction establishes a dynamic regulatory balance that controls cell proliferation and apoptosis, influencing both cellular homeostasis and tumorigenesis.

P73 and p63:

P63 and p73, homologs of p53, play critical roles in epithelial development and contribute to various cancers, though their functions remain less defined compared to p53. These proteins act as tumor suppressors or oncogenes depending on the cellular context and isoform expression [74]. WT1 interacts with all four isoforms of p73 and p63 through its zinc finger domains, modulating their transcriptional activities in a context-dependent manner. These interactions inhibit the

Table 1 WT1 as Oncogene vs Tumor Suppressor

Feature	WT1 as Oncogene	WT1 as Tumor Suppressor
Cancer types	AML, MDS, NSCLC, glioblastoma, ovarian, pancreatic	Wilms' tumor (pediatric kidney cancer), mesothelioma, breast cancer
Expression status	Overexpressed	Loss of function or deletion
Mechanism	Promotes proliferation, inhibits apoptosis	Promotes differentiation, cell cycle arrest, apoptosis
Downstream targets	↑ BCL-2, ↑ VEGF, ↑ MYC, ↑ EGFR	↑ p21, ↑ p53, ↑ IGF-BP, ↑ nephrin, ↑ podocalyxin
Mutation type	Gain-of-function, overexpression	Frameshift, nonsense, or dominant-negative mutations
Therapeutic approach	Target inhibition: vaccines, TCR-T, mAbs, CRISPR	Gene restoration or functional activation
Prognostic value	Poor prognosis if overexpressed	Poor prognosis if inactivated (esp. in syndromic Wilms')
Mouse model phenotype	Overexpression → tumor formation	Knockout → lethal kidney defects



 Table 2
 WT1 interactions with other cancer genes

Aspect	Wilms Tumor I (WT1) [25, 112]	TP53 [72, 73]	KRAS [143]	HER2 (ERBB2)[114]	BCR-ABL[194, 205]
Gene Role	Tumor suppressor (normal) & oncogene (in cancer)	Tumor suppressor	Oncogene	Oncogene	Fusion oncogene
Cancer Types	Leukemia, Wilms tumor, meso- Most solid tumors thelioma, glioblastoma	Most solid tumors	Pancreatic, lung, colon	Breast, gastric	CML, ALL
Mechanism in Cancer	Overexpressed in many cancers; Loss-of-function (LOF) mutaaffects differentiation & tions → loss of cell cycle proliferation control	Loss-of-function (LOF) mutations → loss of cell cycle control	Gain of function (GOF) mutations → uncontrolled growth	Gene amplification → overactive signaling	Fusion protein → constant kinase activation
Targeting Strategies	Vaccines, CAR-T, peptide immunotherapy, antisense, siRNA	Gene therapy, synthetic lethality, reactivating mutants	Direct inhibitors (difficult), KRAS G12C inhibitors	Monoclonal antibodies (trastuzumab), TKIs	TKIs (imatinib, dasatinib)
Therapeutic Challenges	Therapeutic Challenges WT1 has both oncogenic and tumor suppressor roles; nuclear localization limits druggability	Difficult to replace tumor sup- pressor; reactivation should be safe	Targeting mutant protein selectively without affecting normal	Resistance, cardiotoxicity	Resistance mutations
Clinical Trials	Active trials for vaccines (e.g., galinpepimut-S) & CAR-T	Trials on p53 reactivation & gene delivery	KRAS G12C inhibitors (e.g., sotorasib) approved	Many FDA-approved drugs	Highly successful targeted therapy model
Biomarker Use	Yes, for prognosis & MRD in leukemia	Yes, TP53 status affects treatment decisions	Yes, KRAS mutation testing is routine	HER2 testing standard in breast cancer	BCR-ABL monitoring essential



Table 3 WT1 Gene Involvement In Various Type Cancers

Cancer Type	WT1 Involvement	Additional Notes
Wilms' Tumor [12, 27]	Germline and somatic mutations; classic tumor suppressor role	WT1 mutations observed in about 20% of cases; associated with syndromes like WAGR and Denys–Drash
Acute Myeloid Leukemia (AML) [17]	Overexpression; mutations; prognostic marker	High WT1 expression linked to poor prognosis; used as a minimal residual disease marker
Desmoplastic Small Round Cell Tumor [145–147]	EWS-WT1 gene fusion resulting from t(11;22) (p13;q12) translocation	Pathognomonic EWS-WT1 fusion protein drives tumorigenesis
Glioblastoma and Other Gliomas [22, 140–142]	Hyper-expression correlates with tumor grade and proliferation	WT1 expression associated with higher MIB-1 index, indicating increased proliferation
Osteosarcoma [204]	Downregulation of the PI3K/AKT pathway & increased Bax/Bcl2 ratio	WT1 overexpression correlates with aggressive metastatic behavior
Lung Cancer [91, 125]	Overexpression in both small-cell and non- small-cell lung cancers	WT1 expression observed in various lung cancer cell lines and tissues
Colon Cancer [133, 134]	Overexpression in tumor cell lines; variable expression in tissues	WT1 expression detected in colon cancer cell lines; tissue expression varies
Breast Cancer [110–112]	WT1 is shown to influence breast cancer by regulating other genes	Acts as a tumor suppressor or oncogene and affects cancer progression and therapy
Ovarian Cancer [126–128]	Overexpression, particularly in serous carcinoma subtypes	WT1 serves as a diagnostic marker; expression varies among subtypes
Pancreatic Cancer [136–138]	Overexpression in ductal adenocarcinoma	WT1 expression observed in pancreatic cancer tissues
Esophageal Cancer [218]	Overexpression in tumor tissue, enhanced cell migration and invasion	WT1 expression detected in esophageal cancer specimens
Thyroid Cancer [236]	Overexpression in primary thyroid cancers	WT1 expression observed in various thyroid cancer types
Head and Neck Squamous Cell Carcinoma [76]	WT1 regulates p63; promotes proliferation	WT1 involvement in cell proliferation and tumor progression, indicating potential targets for immunotherapy in HNSCC

DNA binding and transcriptional activation abilities of p73 and p63, thereby influencing cell differentiation, apoptosis, and tumor suppression [75]. The complex interplay between WT1 and these p53 family members highlights the intricate regulation of cellular transcriptional networks.

In squamous cell carcinoma of the head and neck (SCCHN), WT1 directly regulates p63 expression and promotes cell proliferation. Silencing WT1 in FaDu cells reduces p63 levels and decreases proliferation, demonstrating that WT1 acts as a positive transcriptional regulator of p63 in this cancer model. Chromatin immunoprecipitation (ChIP) and quantitative PCR analyses confirmed that WT1 binds directly to the promoters of both TAp63 and $\Delta Np63$ isoforms, further supporting its role in transcriptional regulation of p63 [76].

These findings collectively underscore the dynamic regulatory network between WT1 and p53 family proteins in controlling key cellular processes and maintaining genomic integrity. Disruptions in these interactions contribute to tumorigenesis, emphasizing the therapeutic potential of targeting this regulatory axis. Understanding the mechanisms underlying interactions between WT1 and p53 family proteins opens avenues for potential targeted therapies. Modulating these

interactions could restore normal transcriptional regulation, offering potential strategies for treating cancers associated with WT1 and p53 dysfunction.

PTEN

WT1 regulates PTEN expression through multiple mechanisms. In non-small-cell lung cancer (NSCLC), the long non-coding RNA WT1-AS—transcribed from the antisense strand of WT1, acts as a molecular sponge for miR-494-3p. This interaction prevents miR-494-3p-mediated repression of PTEN, thereby inhibiting the PI3K/AKT signaling pathway and suppressing cell proliferation, survival, migration, and invasion. [77]. In Wilms' tumor, bioinformatics analysis identified miR-21-5p as a potential regulator that targets and suppresses PTEN, thereby activating PI3K/AKT pathway and promoting tumorigenesis, including enhanced tumor cell migration and invasion. This finding suggests that altered regulation of PTEN by miRNAs like miR-21-5p could contribute to the pathogenesis of Wilms' tumor [78]. Similarly, increased levels of miR-19b suppress PTEN expression, leading to heightened PI3K/AKT pathway activity and promoting proliferation and apoptosis resistance, while



inhibition of miR-19b reverses these effects [79]. These findings emphasize the multifaceted regulation of PTEN by WT1 and its downstream mediators. Understanding these interactions could offer novel therapeutic opportunities for cancers involving PI3K/AKT dysregulation.

BRCA1

Researchers have not reported any direct interaction between Wilms' Tumor 1 (WT1) and BRCA1. Although both genes function as crucial tumor suppressors involved in DNA repair and cancer progression, their direct interaction remains speculative. Recent studies have further clarified BRCA1's role in chromatin remodeling and DNA damage repair. For example, Witus et al. (2023) demonstrated that intrinsically disordered regions in BRCA1/BARD1 mediate histone H2A ubiquitylation and chromatin recruitment, key processes in the DNA damage response [80]. Wegert et al. (2025) analyzed cases of bilateral Wilms tumor and identified germline variants in WT1, TRIM28, and BRCA2, suggesting possible shared predisposition pathways. However, this study did not implicate BRCA1 in direct functional association with WT1 [81].

In summary, while WT1 and BRCA1 both play significant roles in tumor suppression and DNA repair, future studies are needed to investigate whether any functional relationship exists between WT1 and BRCA1 in cancer biology.

CDC73 (Parafibromin)

WT1 directly represses the tumor suppressor CDC73 in oral squamous cell carcinoma (OSCC) by binding to its promoter and inhibiting its transcription. This repression reduces CDC73 expression, leading to increased cell proliferation and decreased apoptosis. These findings suggest that WT1 functions as an oncogene in this context by actively suppressing the expression of CDC73, a key regulator of transcription and cell cycle control [82].

Bcl-2 family proteins

WT1 directly regulates both pro-apoptotic and anti-apoptotic members of the Bcl-2 family in a context-dependent manner. In some settings, WT1 upregulates anti-apoptotic proteins such as Bcl-2, thereby enhancing cell survival and resistance to apoptosis [83]. In contrast, under different conditions, WT1 activates the expression of pro-apoptotic genes like Bak and Bik, promoting apoptosis [84]. Another study demonstrates that WT1, with Par-4, binds the Bcl-2 promoter and represses its expression, promoting apoptosis [85]. These findings underscore WT1's dual regulatory role in apoptosis, demonstrating that it can either promote

or inhibit cell death by modulating Bcl-2 family proteins depending on the cellular environment.

MAP kinase phosphatase 3 (MKP3)

WT1 induces the expression of MAP kinase phosphatase 3 (MKP3), also known as DUSP6, a negative regulator of the Ras/MAPK signaling pathway. By upregulating MKP3, WT1 decreases ERK phosphorylation, thereby suppressing cell proliferation. This regulation reveals a novel mechanism through which WT1 exerts growth-inhibitory effects by modulating MAPK pathway signaling [86].

β-Catenin

In breast cancer cells, WT1 reduces β -catenin protein levels and impairs β -catenin/TCF transcriptional activity, thereby suppressing Wnt signaling. This destabilization of β -catenin is associated with WT1-mediated inhibition of breast cancer cell growth [87].

These findings underscore the multifaceted role of WT1 in regulating tumor suppressor pathways. The functional outcomes of WT1 interactions vary across cell types and tumor contexts, reflecting its highly context-dependent nature. Further research is needed to fully elucidate the complex network of WT1 interactions with other tumor suppressors and their implications in cancer biology. Despite it initially classified as a tumor suppressor, WT1 also exhibits oncogenic properties in several malignancies, making it a promising target for cancer research and immunotherapy. Therapeutic strategies, such as intracellular antibodies targeting WT1, have shown potential in preclinical studies [88].

Why WT1 is a promising cancer target

Widespread overexpression in cancers

Wilms' Tumor 1 (WT1) serves as a compelling target for cancer immunotherapy due to its widespread overexpression across various hematologic and solid malignancies including leukemia, lung cancer, ovarian cancer, pancreatic cancer, and thymic tumors, while maintaining limited expression in normal adult tissues. Notably, researchers have reported WT1 overexpression in approximately 20% of intrahepatic cholangiocarcinoma (CCA) cases [89, 90].

In non-small-cell lung cancer (NSCLC), Li et al. (2023) demonstrated that WT1 expression is significantly elevated in tumor tissues and cell lines. They further observed that higher WT1 levels correlate with poor prognosis and increased immune cell infiltration, implicating WT1 in NSCLC progression [91]. Collectively, these studies highlight WT1 as a universal tumor-associated antigen. Its aberrant overexpression, association with aggressive disease



features, and limited normal tissue expression underscore its value as a target for therapeutic interventions across a broad spectrum of cancers.

Intracellular localization and immunogenicity

WT1 is an intracellular protein that undergoes antigen processing, allowing tumor cells to present WT1-derived peptides via HLA class I molecules. This presentation enables cytotoxic CD8+ T cells to specifically recognize and eliminate WT1-expressing tumor cells. Immunotherapies, including peptide vaccines, T cell therapies, and CAR-T cells, exploit this mechanism to target WT1-positive cancers. Nishida et al. (2022) showed that WT1 peptide vaccination induces strong cellular and humoral immunity in ovarian cancer, improving clinical outcomes [92]. Liu et al. (2016) showed that WT1 peptide vaccination in myeloid leukemia elicited WT1-specific T-cell responses with TCR clonal enrichment [93]. Further, Liu et al. (2018) demonstrated WT1-specific T-cell responses with TCR clonal enrichment in myeloid leukemia, with Montanide adjuvant enhancing vaccine efficacy [94]. These studies collectively confirm that antigen processing and HLA presentation expose intracellular WT1 to immune recognition, establishing WT1 as a robust and versatile target for cancer immunotherapy.

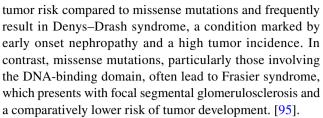
Functional role in tumorigenesis

WT1 promotes tumorigenesis by regulating genes involved in proliferation, survival, and metastasis. In non-small-cell lung cancer (NSCLC), WT1 enhances malignancy by coexpressing with oncogenic factors such as MYC and matrix metalloproteinase 2 (MMP2), which drive cancer progression. Li et al. (2023) demonstrated that microRNA-498-5p suppresses WT1 expression, thereby reducing NSCLC aggressiveness and suggesting a regulatory axis with therapeutic potential [91].

Integrating population studies and WT1-targeted therapies

Integrating population studies with WT1-targeted therapies offers a robust strategy to understand and treat cancers driven by WT1 dysregulation. WT1 functions as either a tumor suppressor or an oncogene, depending on the cancer type and patient-specific genetic context, thereby shaping therapeutic decisions.

Population-based studies have defined the clinical relevance of specific WT1 mutations in determining cancer risk and prognosis. Nagano and Nozu (2025) analyze genotype-phenotype correlations in WT1-related disorders, including Wilms tumor. Their review highlights that truncating mutations in WT1 associate with a higher Wilms



The study emphasizes the importance of understanding these genetic variations to guide personalized treatment strategies. By elucidating genotype-phenotype relationships, they promote the development of mutation-specific therapeutic approaches. Clinicians can use this mutation-specific information to predict disease progression and tailor interventions accordingly. Previous study analyzed WT1 exon 8 and 9 missense mutations in the DNA-binding domain and demonstrated that these mutations produce diverse phenotypes, ranging from mild nephropathy to varying tumor risks. The findings confirmed that missense mutations generally cause milder clinical symptoms than truncating mutations and underscore the importance of detailed genetic analysis to guide personalized treatment of WT1-related disorders [96]. Similarly, Lipska et al. (2014) investigated 64 patients with WT1-related glomerulopathy and identified clear genotype-phenotype correlations. They found that dominant-negative missense mutations in the zinc finger domains typically led to Frasier syndrome, which involves slow-progressing kidney disease and gonadal dysgenesis. In contrast, truncating mutations or those outside the zinc finger domains often resulted in Denys-Drash syndrome, which presents with early onset nephrotic syndrome and increased Wilms tumor risk. Their study confirmed that the type and location of WT1 mutations significantly influence disease severity and progression [97]. Liu et al. (2023) analyzed WT1 mutations in Chinese children with nephropathy and further supported the genotype-phenotype correlations described by Nagano and Nozu. They found that truncating mutations cause severe early onset nephropathy and increase tumor risk, while missense mutations lead to milder phenotypes. This population-specific data highlight the importance of integrating genetic insights to guide personalized treatment approaches for WT1-related disorders [98].

In conclusion, integrating comprehensive population-level genetic data with emerging WT1-targeted therapeutic approaches provides a powerful framework to enhance our understanding of WT1's dual role as a tumor suppressor and oncogene across diverse cancer types. Insights from genotype-phenotype correlation studies enable precise identification of mutation-specific risks and clinical outcomes, which are critical for tailoring personalized treatment strategies. Building on this foundation, innovative therapies such as antisense oligonucleotides (ASOs) [99] and small interfering RNAs (siRNAs) [100–102] offer promising tools to selectively inhibit oncogenic WT1 isoforms in both hematologic



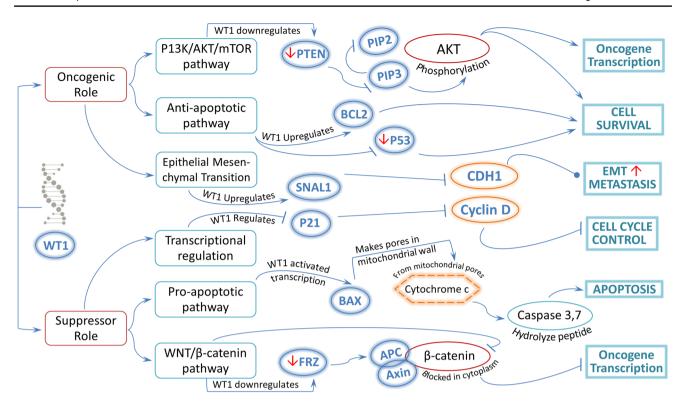


Fig. 2 The Role of WT1 in Cancer Pathogenesis through the Regulation of Target Genes: The Wilms tumor 1 (WT1) gene is primarily recognized as a tumor suppressor, acting as a transcriptional repressor; however, in certain cancers, it can function as an oncogene by serving as a transcriptional activator [42]. The biological effects of WT1 expression are context-dependent, influenced by the specific cell type and the surrounding regulatory microenvironment, which collectively determine whether WT1 promotes cellular proliferation or exerts growth-inhibitory effects. As a tumor suppressor, WT1 regulates transcriptional pathways such as the activation of p21, which inhibits Cyclin D and controls cell cycle progression. WT1 also pro-

motes apoptosis through the activation of the pro-apoptotic protein BAX. Additionally, WT1 suppresses the Wnt/β-catenin signaling pathway, further contributing to its tumor suppressive functions [84, 87]. Conversely, when WT1 acts as an oncogene, it promotes tumor invasion and metastasis through epithelial–mesenchymal transition (EMT), partly by repressing E-cadherin expression [127]. WT1 also enhances the transcription of Bcl-2, an anti-apoptotic protein that supports cell survival [83]. Furthermore, WT1 downregulates the tumor suppressor gene PTEN, inhibiting its phosphatase activity and preventing the dephosphorylation of PIP3, thus contributing to cellular growth and survival [78]

malignancies and solid tumors. Concurrently, immunotherapeutic interventions including WT1 peptide vaccines are being actively developed and clinically evaluated to stimulate targeted anti-tumor immune responses [103, 104]. By leveraging population genetics alongside molecularly informed WT1-targeted treatments, clinicians and researchers can advance mutation-guided precision oncology that maximizes therapeutic efficacy while minimizing adverse effects, ultimately improving prognosis and quality of life for patients with WT1-associated cancers.

Identification of Wilms' tumor 1 (WT1) target genes and its role in cancer pathogenesis through the regulation of other genes

WT1 regulates numerous genes involved in critical cellular processes, including proliferation, differentiation, apoptosis, survival, and metastasis. Acting as a transcriptional regulator, WT1 either activates or represses these target genes,

thereby driving diverse biological outcomes such as cell growth, differentiation, and apoptosis (Fig. 2). Researchers have identified a wide array of WT1 target genes through microarray analyses [105–107]. These target genes include AREG, CDKN1A (p21/CIP1), DAX1, colony-stimulating factor 1 (CSF-1), Müllerian-inhibiting substance (MIS), SF1, sex-determining region Y (SRY), CDH1 (E-cadherin), podocalyxin, nephrin (NPH1), BAK, and NTRK2. Together, they mediate the diverse regulatory functions of WT1 in essential cellular processes [84, 108, 109].

A genome-wide expression profiling study identified several direct target genes of WT1, including members of the EGF family of growth factors such as AREG, EREG, and HB-EGF, along with the chemokine CX3CL1 and the cytokine IL-11. Additional analyses using qRT-PCR, chromatin immunoprecipitation (ChIP), and immunohistochemistry further confirmed other potential direct targets of WT1, including IL1RAP, SLUG, JUNB, SLC20A1, and TIMP3 [40].



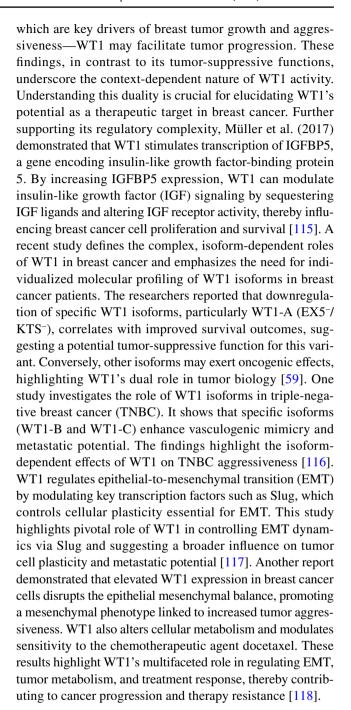
We present a concise overview of the role of WT1 in various cancers, emphasizing its regulatory impact on gene expression and underscoring its clinical relevance as both a prognostic marker and a potential therapeutic target across multiple cancer types.

Breast cancer

Elevated WT1 expression is commonly observed in breast cancer cells; high WT1 mRNA expression is associated with poorer disease-free survival. While no significant correlation was found between WT1 levels and clinicopathological parameters such as tumor size or lymph node status, the prognostic value of WT1 expression suggests its potential utility as an independent prognostic biomarker [110]. Recent study reports that WT1 is overexpressed in triple-negative breast cancer (TNBC), correlates with poor prognosis, and drives metastasis by transcriptionally activating PFKFB4 to enhance glycolysis. Co-expression of WT1 and PFKFB4 is linked to worse clinical outcomes, highlighting WT1's oncogenic and prognostic significance in TNBC [111].WT1 regulates genes involved in proliferation, migration, apoptosis, and survival, influencing key oncogenic and tumor suppressor pathways such as p53, EphA2, HER2, ER-α, and TPα. It controls caspase family members that mediate apoptosis and affect tumor progression. In hormone-responsive breast cancer, WT1 modulates estrogen receptor signaling by regulating genes in estrogen biosynthesis and cellular response to estrogen. These roles establish WT1 as a critical factor in breast cancer pathophysiology and a promising prognostic biomarker and therapeutic target [112].

Previous studies have suggested that WT1 may function as a tumor suppressor gene in breast cancer by inhibiting cell proliferation. For instance, one study reported that WT1 suppresses breast cancer cell growth by destabilizing β-catenin, thereby inhibiting the Wnt/β-catenin signaling pathway, which promotes tumor progression [87]. Additionally, WT1 plays a complex and context-dependent role through its interactions with estrogen receptor alpha (ER α) and growth factor signaling pathways. Reizner et al. (2005) demonstrated that WT1 interacts with estrogen receptor alpha (ERα) and inhibits its transactivation of the insulinlike growth factor 1 receptor (IGF-1R) gene in breast cancer cells. This inhibition reduces IGF-1R expression and subsequently suppresses cell proliferation, further supporting WT1's tumor-suppressive function in specific breast cancer contexts. [113].

Conversely, Nasomyon et al. (2014) reported that WT1 promotes breast cancer progression by upregulating the expression of ER α , human epidermal growth factor receptor 2 (HER2), and epidermal growth factor receptor (EGFR), suggesting an oncogenic role for WT1 in certain cellular contexts [114]. By enhancing ER α and HER2 expression,



Acute myeloid leukemia (AML)

In acute myeloid leukemia (AML) and myelodysplastic syndromes (MDS), WT1 is frequently overexpressed and occasionally mutated, highlighting its role in leukemogenesis and potential as a therapeutic target. Approximately 70–90% of AML patients show WT1 overexpression, while 6–15% carries WT1 mutations [119]. These mutations frequently occur in conjunction with other genetic alterations, such as FLT3-ITD and NPM1 mutations, contributing to disease aggressiveness and promote leukemogenesis. WT1 also modulates



several molecular pathways in leukemic cells, supporting the survival and self-renewal of leukemia-initiating cells (LICs) [120–122]. It also interacts with epigenetic regulators like DNMT3A, further promoting cancer cell growth and survival in AML patient [123].

Myelodysplastic SYNDROMES (MDS)

WT1 is frequently overexpressed in MDS and correlates with poor overall survival (OS). However, emerging evidence suggests that the prognostic relevance of WT1 expression in myelodysplastic syndromes (MDS) is modulated by the presence of specific genetic mutations, notably alterations in TP53. These findings underscore the multifaceted role of WT1 in MDS pathogenesis and highlight the necessity of integrating mutational profiling particularly TP53 status into prognostic stratification frameworks. [124].

Non-Small-Cell lung cancer (NSCLC)

WT1 is upregulated in NSCLC tissues and cell lines and correlates with poor prognosis and increased immune cell infiltration. Functional assays show that WT1 promotes cell proliferation, survival, and invasion. Survival analyses indicate that elevated WT1 expressions correlate with decreased overall and progression-free survival, supporting its role as a prognostic biomarker and therapeutic target [91]. Micro-RNA-498-5p acts as a natural inhibitor of WT1, indicating a potential therapeutic strategy to suppress WT1 expression in NSCLC. Xu et al. (2013) further demonstrated that WT1 is overexpressed in NSCLC tissues relative to adjacent normal tissues and its expression positively correlates with cell proliferation in both cell lines and a xenograft mouse model. This study revealed that WT1 enhances cell proliferation by upregulating Cyclin D1 and phosphorylated retinoblastoma protein (p-pRb), facilitating S-phase entry and cell cycle progression. These findings establish WT1 as an oncogenic driver in NSCLC through modulation of cell cycle regulators [125].

Ovarian carcinoma

WT1 plays a significant role in ovarian cancer. Liu et al. (2014) reported that WT1 overexpression correlates with aggressive clinical features such as advanced stage, poor differentiation, and reduced overall survival, suggesting WT1 as a biomarker for tumor aggressiveness and prognosis. [19]. In contrast, Taube et al. (2016) found that WT1 is commonly expressed in high-grade serous ovarian carcinoma (HGSC) and associates with improved overall survival (OS) and progression-free survival (PFS), independent of age, stage, and residual tumor status [126]. Han et al. (2020) demonstrated

that WT1 promotes ovarian cancer progression by regulating E-cadherin and ERK1/2 signaling. WT1 directly binds the E-cadherin promoter, suppressing its expression and inducing epithelial-to-mesenchymal transition (EMT) via ERK1/2 signaling pathway activation, thus driving tumor invasiveness. Targeting WT1 could inhibit these oncogenic pathways and offer therapeutic benefits [127]. Van Amerongen et al. (2022) identified novel WT1-derived peptides and engineered WT1-specific T cell receptors (TCRs) that selectively recognize and target WT1-expressing tumor cells. These TCRs exhibit potent antitumor activity against acute myeloid leukemia (AML) and ovarian carcinoma in preclinical models, supporting WT1-targeted TCR immunotherapy as a promising treatment for WT1-positive malignancies [128].

Kidney Wilms tumor

WT1 regulates several genes implicated in Wilms tumor (WT) and fetal kidney development. Zeng et al. (2024) identified EMCN and CCNA1 as key hub genes, demonstrating that EMCN overexpression and CCNA1 knockdown significantly reduced Wilms tumor cell viability, proliferation, migration, and invasion, indicating their roles as potential prognostic biomarkers and therapeutic targets. [129]. WT1 also governs genes involved in mesenchymal-to-epithelial transition (MET), a process essential for nephrogenesis. In WT1-mutant Wilms tumors, concurrent CTNNB1 mutations frequently activate Wnt/ β-catenin signaling, which disrupts epithelial differentiation and promotes abnormal stromal cell proliferation. This molecular interplay impairs kidney architecture and promotes tumorigenesis [130]. Royer-Pokora et al. (2023) further demonstrated that WT1-mutant Wilms tumors exhibit gene expression profiles aligned with early renal stem cell states, supporting their origin from undifferentiated nephron progenitors [131]. In parallel, Rialdi et al. (2023) showed that WNTinib, a selective multi-kinase inhibitor, effectively blocks Wnt/β-catenin signaling in β-catenin-mutant tumors. Given the frequent activation of β-catenin in WT1-mutant Wilms tumors, WNTinib may offer a promising targeted therapeutic strategy in this context [132]. These findings support WT1 as a diagnostic and prognostic marker and emphasize the therapeutic potential of targeting the Wnt/β-catenin pathway in Wilms tumor.

Renal cell carcinoma (RCC)

WT1 exhibits tumor-suppressive activity in renal cell carcinoma (RCC). Jing et al. (2022) demonstrated that WT1 overexpression in human RCC A498 cells significantly



336

upregulates interleukin-24 (IL-24), a cytokine with antitumor properties. This induction leads to G2/M cell cycle arrest and marked inhibition of cell proliferation, indicating a direct role of the WT1/IL-24 axis in tumor suppression. Furthermore, analysis of The Cancer Genome Atlas (TCGA) data revealed that WT1-regulated genes such as TXNIP and GADD45A correlate inversely with tumor stage and histological grade. Elevated expression of these genes associates with favorable prognoses, reinforcing WT1's protective function in RCC progression [16].

These findings underscore WT1's context-dependent role in tumor biology and support the WT1/IL-24 signaling pathway as a promising therapeutic target, particularly in RCC cases, especially for those resistant to conventional therapies [133].

Endometrial carcinoma

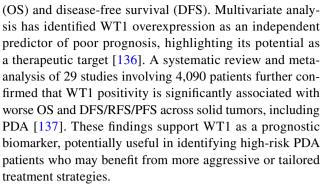
In endometrial carcinoma, especially the serous and clear cell subtypes, elevated WT1 expression correlates with poorer overall survival and disease-free survival. WT1 levels associate positively with advanced tumor stage and higher histological grade, highlighting its potential as a prognostic marker [134]. However, studies have yet to establish WT1 as an independent prognostic factor, and further validation is required. Current studies actively explore WT1-targeted therapies to develop personalized treatment strategies in endometrial carcinoma [133].

Uterine serous carcinoma (USC)

WT1 plays a potential diagnostic and prognostic role in uterine serous carcinoma (USC), where its expression may influence treatment outcomes. WT1 expression is detected in over 20% of uterine serous carcinoma (USC) cases [135]. Notably, WT1-positive tumors exhibit significantly greater chemosensitivity, which correlates with a trend toward improved progression-free survival [135]. Another study concluded that WT1 expression correlates with poor prognosis in certain histotypes and may serve as both a diagnostic and prognostic marker, particularly in high-grade subtypes like USC. Evaluating WT1 status in USC could help predict chemotherapy response, stratify patients by risk and guide personalized treatment strategies [134].

Pancreatic ductal adenocarcinoma (PDA)

In pancreatic ductal adenocarcinoma (PDA), WT1 is predominantly expressed in the cytoplasm of tumor cells and within tumor-associated vasculature. Higher cytoplasmic WT1 expression correlates with shorter overall survival



In addition, WT1-targeted immunotherapy, such as peptide-loaded dendritic cell vaccines, has shown promise in PDA, especially when combined with standard chemotherapy, indicating WT1's relevance as a target for novel immunotherapeutic interventions [138].

Soft tissue sarcomas (sts)

WT1 is frequently expressed in several soft tissue sarcoma (STS) subtypes, including rhabdomyosarcoma and malignant peripheral nerve sheath tumors. Interestingly, in highgrade STS, strong WT1 expression correlates with improved survival outcomes. This paradoxical finding underscores its context-dependent function in tumor biology and highlights the need for further research to understand its prognostic significance and potential for immunotherapy. [139].

Glioblastoma and other glial tumors

WT1 protein is expressed in nearly all glioblastomas and anaplastic glial tumors. Studies show a significant positive correlation between WT1 expression and cellular proliferation markers, such as the MIB-1 index, indicating WT1's role in promoting tumor cell proliferation. These findings support WT1 as a potential therapeutic target in malignant glial tumors [140–142].

KRAS-driven cancers

In KRAS-driven cancers, WT1 modulates oncogenic signaling and cellular senescence. WT1 interacts with KRAS, a key regulator of proliferation and survival, and exhibits context-dependent roles—either promoting tumor growth or inducing senescence. In KRAS-driven lung cancer models, WT1 loss induces senescence, suggesting a tumor-suppressive function [143]. This effect is mediated by WT1's regulation of metabolic and oncogenic pathways, including MYC and glutamine metabolism, which are critical for tumor maintenance.

Disruption of WT1 impairs these pathways, reducing cell viability in KRAS-mutant tumors. Additionally, WT1 serves as a potential predictive biomarker for immunotherapy



response in non-small-cell lung cancer (NSCLC), linking its role to immune modulation and prognosis [144]. These findings position WT1 as a dual-function therapeutic target in KRAS-driven malignancies—either to enhance senescence or suppress oncogenic activity.

Desmoplastic small round cell tumor (dsrct)

The WT1 gene, particularly as part of the EWSR1::WT1 fusion oncogene, drives pathogenesis of desmoplastic small round cell tumor (DSRCT) by regulating key pathways, including upregulating CCND1, which promotes tumor proliferation via the cyclin D–CDK4/6–RB axis. Molecular profiling of DSRCTs also reveals overexpression of growth factor receptors such as FGFR4, IGF2, and ERBB2, identifying additional therapeutic targets. These molecular insights highlight CDK4/6 inhibitors and other targeted therapies as promising strategies for desmoplastic small round cell tumors [145–147].

Collectively, the evidence underscores WT1 as a pivotal regulator of tumor biology across a wide spectrum of malignancies. Its regulatory roles span key oncogenic processes, including gene transcription, apoptosis, cell proliferation, epithelial-to-mesenchymal transition (EMT), immune evasion, and metabolic reprogramming.

WT1 overexpression is consistently associated with adverse clinical outcomes in several cancers such as breast cancer, AML, NSCLC, and pancreatic ductal adenocarcinoma, highlighting its utility as a negative prognostic biomarker. Conversely, in select tumors like renal cell carcinoma and certain soft tissue sarcomas, WT1 exhibits tumor-suppressive activity, further reinforcing its functional complexity. Isoform-specific effects in cancers such as breast cancer and Wilms tumor emphasize the importance of precise molecular characterization of WT1 variants for prognostic and therapeutic purposes.

Wilms' tumor 1 (WT1) gene as a marker for minimal residual disease (MRD)

Minimal residual disease (MRD) comprises small populations of malignant cells that persist after treatment and evade detection by conventional diagnostics such as imaging and standard laboratory tests. These residual cells can lead to disease relapse, particularly in hematologic malignancies including leukemia, lymphoma, and multiple myeloma. Recent studies underscore the prognostic and therapeutic significance of MRD monitoring, which complements baseline clinical, cytogenetic, and molecular data. MRD assessment enables early relapse prediction and informs treatment decisions, especially in post-transplant settings. Its utility is well established in acute lymphoblastic leukemia (ALL),

acute promyelocytic leukemia (APL), and chronic myeloid leukemia (CML) [148–151].

The Wilms' Tumor 1 (WT1) gene has emerged as a useful molecular marker for measurable residual disease (MRD) monitoring in acute myeloid leukemia (AML), particularly in patients lacking specific genetic aberrations [152]. Although WT1 is not AML-specific and is classified as a pan-leukemic marker, it is overexpressed in various leukemias and occasionally detected in normal bone marrow [153–155]. Its expression in peripheral blood allows for noninvasive monitoring of disease burden [156]. Cilloni et al., in collaboration with the European LeukemiaNet (ELN), developed a standardized qRT-PCR assay for reproducible quantification of WT1 transcripts in bone marrow and peripheral blood samples [157, 158]. The ELN recommends WT1 thresholds of 250 copies per 10⁴ ABL copies in bone marrow and 50 copies per 10⁴ ABL copies in peripheral blood to distinguish residual disease from normal expression, optimizing sensitivity and specificity [159].

Although WT1 is widely used as an MRD marker, its expression also occurs in healthy individuals, which can complicate interpretation. To reduce false positives, clinicians should interpret WT1 results alongside clinical data and complementary MRD tools like flow cytometry and genomic analyses. Advances in PCR methodologies and standardized guidelines have enhanced WT1 MRD detection reliability. Next-generation sequencing (NGS) represents a promising, though still evolving, MRD surveillance tool [160].

As a diagnostic and prognostic marker

The Wilms' Tumor 1 (WT1) gene is overexpressed in over 80% of acute myeloid leukemia (AML) cases, making it a broadly applicable marker for measurable residual disease (MRD) detection, particularly in patients lacking mutations such as NPM1 or fusion genes like RUNX1-RUNX1T1 [159, 161]. The WT1 overexpression is utilized as a diagnostic biomarker, while WT1 expression levels predict outcomes and fulfilling a prognostic marker rule. Studies have consistently demonstrated significantly elevated WT1 expression in AML patients compared to healthy controls, supporting its role in MRD monitoring and outcome prediction, and revealing that WT1 mRNA as a reliable prognostic marker for treatment response and relapse risk in AML [151, 162] The paper by Ugale et al. (2024) investigates the prognostic impact of WT1 gene mutations in acute myeloid leukemia (AML). It demonstrates that WT1 mutations are associated with poor risk stratification and reduced overall survival. The study supports incorporating WT1 mutation status into AML prognostic models for better clinical outcomes. [163]. One study examines the expression of Wilms tumor gene 1 (WT1) in high-grade uterine sarcomas. It demonstrates



336

that WT1 is frequently expressed in these tumors and is associated with poor prognosis. The study suggests WT1 as a potential prognostic biomarker and therapeutic target in high-grade uterine sarcoma [164]. In non-small-cell lung cancer (NSCLC), WT1 promotes tumor malignancy and serves as a novel prognostic biomarker. Inhibition of WT1 by microRNA-498-5p reduces tumor aggressiveness, indicating WT1 as a potential therapeutic target [91]. These findings establish WT1 as a key prognostic marker for guiding personalized cancer treatment.

Post-Treatment monitoring

Multiple studies demonstrate that WT1 plays a critical role in post-treatment monitoring of AML by tracking remission status and predicting relapse. Detectable WT1 mRNA in patients who achieve complete remission correlates with a higher relapse risk. WT1's high sensitivity allows detection of residual leukemic cells even when cytogenetic or other molecular markers are undetectable [158, 159, 162].

Prognostic marker for Post-Hematopoietic stem cell transplantation (HSCT)

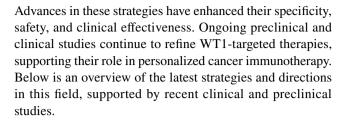
WT1 serves as a critical prognostic marker after hematopoietic stem cell transplantation (HSCT), with elevated WT1 levels early post-transplant strongly predicting disease relapse. In such cases, clinicians may initiate donor lymphocyte infusion or preemptive chemotherapy to mitigate relapse risk [158, 165].

In allogeneic stem cell transplantation (allo-SCT) patients, increased WT1 expression correlates with higher relapse incidence. Combining WT1 monitoring with other MRD detection methods, such as flow cytometry, improves relapse prediction and guides post-transplant treatment decisions [166].

WT1: A therapeutic target for different approaches of cancer treatment

WT1 as a Tumor-associated antigen

Wilms' tumor 1 (WT1) is overexpressed in numerous hematologic and solid malignancies and has been identified as a prominent tumor-associated antigen (TAA). Its selective expression in cancer cells compared to normal tissues makes WT1 an ideal target for immunotherapy. WT1-derived peptides induce robust cytotoxic T lymphocyte (CTL) and helper T cell responses that specifically eliminate WT1-expressing tumor cells while sparing healthy cells [103, 104]. Current therapeutic approaches include peptide vaccines, T-cell therapies, and gene-silencing techniques designed to maximize antitumor efficacy and minimize off-target toxicity (Fig. 3).



Potential strategies in WT1-directed therapies

wt1-based immunotherapeutic vaccines

WT1 peptide vaccines activate the immune system to specifically recognize and eliminate WT1-expressing tumor cells. These vaccines deliver WT1-derived peptides, which antigen-presenting cells present via major histocompatibility complex (MHC) molecules, thereby stimulating WT1-specific cytotoxic T lymphocytes (CTLs) to target and destroy tumor cells expressing WT1 (Fig. 4). Clinical studies demonstrate that WT1 vaccines safely induce strong immune responses across various tumor types. Combining HLA class I and II peptides enhances these responses, correlating with improved clinical outcomes and prolonged survival, especially in hematologic malignancies [103, 104, 167–170]. A randomized Phase II trial in elderly AML patients showed that immunoreactivity to the WT1 peptide vaccine (OCV-501) correlated with improved prognosis, supporting its therapeutic potential in this patient population [171]. In pediatric oncology, WT1-targeted vaccination showed clinical benefit in children with relapsed rhabdomyosarcoma and diffuse midline glioma, leading to stable disease and improved outcomes. These findings underscore the potential of WT1-targeted therapies in pediatric refractory cancers [172]. A Phase I clinical study investigated a cocktail vaccine of WT1 HLA class I and II peptides for recurrent malignant glioma. The study demonstrated the feasibility and safety of this approach, with indications of immune responses, highlighting its potential as a therapeutic strategy for glioma patients [173]. Recent advancements include WT1 peptide-pulsed dendritic cell (WT1-DC) vaccines and WT1 mRNA-electroporated dendritic cell (WT1 mRNA-DC) vaccines, both of which significantly boost antitumor immunity in diverse cancers [104].

Preclinical and Clinical Evidence Supporting WT1 Vaccine Efficacy

Wilms' tumor 1 (WT1) is a well-characterized tumor-associated antigen widely expressed in hematologic and solid malignancies. WT1-based immunotherapeutic vaccines have shown promising preclinical and clinical outcomes.



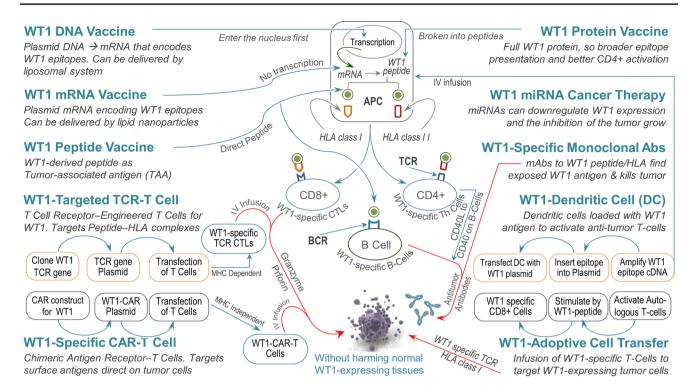


Fig. 3 This figure presents an integrated summary of thirteen representative therapeutic strategies targeting the WT1 gene in cancer, emphasizing its dual role as an oncogene and an immunogenic target. Each approach is described at the molecular or cellular level, beginning with WT1 DNA vaccines [190, 191], which activate cytotoxic T lymphocytes against WT1-expressing tumors and stimulate helper T cell responses. Cellular immunotherapies, including dendritic cell (DC) vaccines [68] and T cell receptor-engineered T cell (TCR-T) therapies [200], enhance tumor-specific immune activation. Monoclonal antibodies [104, 194] and bispecific T cell engagers [196, 197] either disrupt WT1-associated signaling pathways or redirect immune

effector cells to tumor sites. The figure also presents RNA-based therapies, such as WT1-targeting siRNAs [205, 206] and antisense oligonucleotides, which suppress WT1 expression at the transcriptional or post-transcriptional level. Additionally, CRISPR/Cas9 gene editing [202] offers permanent genomic disruption of oncogenic WT1 alleles. Small molecule inhibitors target downstream effectors of WT1, while epigenetic modulators reverse aberrant gene expression patterns driven by WT1. Collectively, these strategies illustrate how WT1's biological functions are being leveraged across diverse therapeutic platforms to treat both hematologic and solid tumors

A systematic review published in Cureus highlighted the application of the WT1 vaccine in advanced pancreatic cancer, emphasizing its potential to improve clinical outcomes by inducing specific immune responses targeting tumor cells [174].

A significant Phase I/II study evaluated the use of a WT1 recombinant protein vaccine in elderly patients with acute myeloid leukemia (AML) in remission. The study demonstrated the vaccine's ability to stimulate both humoral and cellular immune responses, with detailed analysis of minimal residual disease (MRD) and WT1 expression providing further insights into its therapeutic efficacy [175].

Suwabe et al. (2024) reported durable WT1-specific CD8⁺ CTL responses lasting up to ten years following WT1 peptide vaccination, underscoring the vaccine's role in sustaining long-term remission and preventing cancer recurrence [176].

Dendritic cell-based WT1 vaccines and immune activation Dendritic cells (DCs) loaded with WT1 peptides or mRNA function as potent antigen-presenting cells that elicit strong immune responses by bypassing HLA restrictions and activating both CD4⁺ helper and CD8⁺ cytotoxic T cells [177].

Recent studies demonstrate that WT1-mRNA-loaded DC vaccines induce type 1 CD4⁺ and CD8⁺ T-cell responses, achieving disease control and prolonged survival in patients with advanced solid tumors such as glioblastoma, metastatic breast cancer, and malignant pleural mesothelioma. These advances emphasize WT1-targeted immunotherapy's growing role, particularly in relapsed or refractory acute myeloid leukemia [177, 178].

Berneman et al. showed WT1-mRNA dendritic cell vaccines elicit strong type 1 T-cell responses in solid tumor patients, correlating with improved clinical outcomes. The study supports the therapeutic potential of WT1-targeted dendritic cell vaccines in eliciting antitumor immunity [68]. Despite their promise, DC vaccines require complex manufacturing and often predominantly stimulate CD4+ T cells, which may limit effective cytotoxic T lymphocyte (CTL)



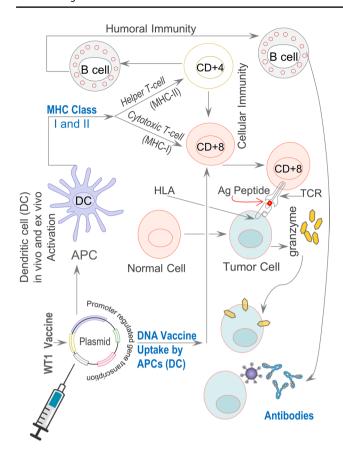
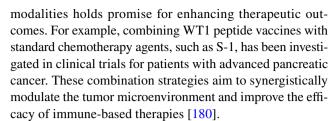


Fig. 4 The WT1 vaccine initiates an immune response that results in the destruction of tumor cells: Oncogenic factors continuously drive the malignant transformation of cells, but the immune system typically identifies and eliminates most of these transformed cells. DNA vaccines, such as the WT1 vaccine, stimulate innate immune cellsincluding dendritic cells (DCs), natural killer (NK) cells, and natural killer T (NKT) cells—that are capable of recognizing and eliminating transformed cells. Antigens captured by antigen-presenting cells (APCs), such as dendritic cells, presented to other immune cells, initiating adaptive immune responses mediated by the WT1 vaccine to target and attack cancer cells. The WT1 vaccine enhances the immune system's ability to recognize tumor-specific antigens and activates both T cells and B cells to mount a targeted immune response against the cancer, without inducing significant autoimmune side effects, thereby facilitating a successful and effective clinical outcome [104, 190]

responses. IL-15-transfected DCs improve T-cell activation and facilitate scalable batch production, offering a significant advancement in vaccine design [179].

Combination therapies to enhance WT1-directed immunotherapy

Recent advancements have concentrated on improving the immunogenicity and therapeutic efficacy of WT1-directed immunotherapies through the use of other treatment modalities such as adjuvants and combination therapies. Combining WT1-directed immunotherapies with other treatment



Preclinical studies demonstrate that combining WT1 vaccines with therapies such as checkpoint inhibitors and chemotherapy synergistically enhances immune responses and overcomes tumor immune evasion. Manning-Geist et al. (2023) conducted a Phase I trial evaluating the multivalent WT1 peptide vaccine galinpepimut-S combined with the checkpoint inhibitor nivolumab in patients with WT1expressing ovarian cancer in second or third remission. The combination induced WT1-specific T-cell responses in 91% of patients and achieved a 70% progression-free survival rate at one year, highlighting its potential to augment antitumor immunity and improve clinical outcomes [181]. The study P4.06 reports that combining WT1 peptide vaccination with imatinib therapy in a patient with chronic myeloid leukemia (CML) led to a sustained long-term molecular remission. This remission correlated with an increased frequency of WT1-specific cytotoxic T lymphocytes (CTLs) even after stopping the WT1 vaccine, suggesting the combination enhances durable immune control of the disease [182].

A phase I trial investigated the use of granulocyte-macrophage colony-stimulating factor (GM-CSF) or CpG oligodeoxynucleotides (CpG-ODN) as immunoenhancement adjuvants in Wilms' Tumor 1 (WT1) peptide vaccine therapy for patients with solid malignancy. The study found that the addition of CpG-ODN improved the effectiveness of the vaccine, with a disease control rate of 60% in the CpG-ODN group [183]. The choice of adjuvants plays a critical role to boost immune responses to WT1 peptide vaccines. For instance, the combination of Montanide ISA-51 and CpG7909 has been used to enhance immune responses. However, studies have shown that suboptimal concentrations of these adjuvants can impair the activation of antigen-presenting cells and the subsequent T-cell responses. Therefore, optimizing adjuvant formulations is essential for improving vaccine outcomes [184]. One notable study investigated an oral WT1 protein vaccine using Bifidobacterium longum 420, to deliver an oral WT1 protein vaccine in leukemiabearing mice, which induced robust WT1-specific cytotoxic T lymphocyte (CTL) responses and activated helper T cells. This vaccine elicited intestinal immunity, enhancing antitumor activity via WT1-specific CD8+ T cells supported by CD4⁺ T cell help with reduced tumor volume correlating to increased CTL frequency, demonstrating potential for systemic and local immune activation against WT1-expressing tumors [185].



Minagawa et al. (2023) developed a combination oral WT1 vaccine using B. longum 420 and 2656, containing a helper epitope. This formulation amplified CD4⁺ T cell-mediated immune responses and enhanced antitumor efficacy in the intestinal tract of leukemia models by promoting WT1-specific immune cell activation [186].

Another study developed a WT1-pulsed dendritic cell vaccine using human CD141⁺ DCs matured with zoledronate, enhancing vaccine efficacy. This approach robustly activated CD8⁺ T cells and elicited strong cytotoxic T lymphocyte (CTL) responses against solid tumors [187].

A Phase II randomized study evaluated the combination of gemcitabine chemotherapy and WT1 peptide vaccination in patients with advanced pancreatic ductal adenocarcinoma (PDAC). The combination significantly improved progression-free survival, particularly in patients who mounted a strong WT1-specific cytotoxic T lymphocyte response. These findings support the potential of combining chemotherapy with WT1-targeted immunotherapy to enhance clinical outcomes in PDAC [188] A Phase I study evaluated the safety and immunogenicity of recombinant WT1 protein combined with the immunostimulant AS15 as neoadjuvant therapy in WT1-positive breast cancer patients. The study found that the combination was well-tolerated and induced WT1-specific antibodies, particularly in patients receiving neoadjuvant aromatase inhibitors [189].

Collectively, these studies suggest that WT1-targeted vaccines, especially when combined with other immunotherapies, hold substantial promise for the treatment of various cancers by effectively stimulating the immune system to recognize and eliminate WT1-expressing tumor cells.

Personalized neoantigen vaccines

Because WT1 functions as a self-antigen, the immune system tightly regulates its expression to maintain tolerance. To overcome this, researchers now design personalized vaccines that target tumor-specific neoantigens—mutated self-antigens that the immune system can recognize as foreign. These vaccines use next-generation sequencing to identify neoantigen targets, including WT1-derived peptides, and trigger strong, antigen-specific T cell responses. Zhang et al. confirmed the safety and immunogenicity of DNA-based neoantigen vaccines in triple-negative breast cancer, demonstrating clonal T cell expansion through single-cell TCR sequencing. This clinical evidence supports the use of neoantigen-targeted immunotherapy as a promising approach in cancer treatment [190, 191].

WT1-Specific antibodies/wt1-specific antibody-based therapies

Monoclonal antibodies

Monoclonal antibodies are engineered proteins that recognize and bind specific antigens on target cells. When targeting WT1, they selectively deliver cytotoxic agents to WT1-expressing tumor cells, minimizing off-target toxicity. However, still in early stage development, this strategy offers a promising approach for precise cancer therapy [104].

WT1-Specific T cell engagers (e.g., WT1-TOPAbody)

Recent advancements in the use of WT1-specific antibodies in cancer therapy include the WT1-specific T cell engagers, such as WT1-TOPAbody, which exhibit high affinity and specificity for the WT1/HLA-A*02:01 complex. This tri-specific construct activates T cells through CD3 and 4-1BB signaling, inducing potent anti-tumor responses in both hematologic malignancies and solid tumors. Preclinical studies have demonstrated its efficacy in WT1/HLA-A02+cancers, including leukemia, ovarian, and breast cancers [192, 193].

TCR-Mimic (TCRm) antibodies

TCR-mimic antibodies developed using Biocytogen's platform target intracellular WT1 peptides presented by MHC class I molecules. These antibodies bind with high specificity to the WT1/HLA complex, enabling selective recognition and destruction of WT1-expressing tumor cells. Preclinical models have confirmed their therapeutic potential in acute myeloid leukemia (AML), ovarian cancer, and other WT1-positive tumors [100, 193, 194]. Another study reports TCR-mimic antibody—drug conjugates targeting WT1/HLA-A*02, showing selective killing of WT1-positive tumor cells and strong antitumor effects in models [195].

Bispecific antibodies (e.g., WT1-TCB)

Novel bispecific antibodies, such as the RMF-peptide-MHC-specific T-cell bispecific antibody (WT1-TCB), have been developed to redirect T cells to WT1-expressing tumor cells. In preclinical models, WT1-TCB induced dose-dependent antitumor activity, effectively targeting leukemia stem cells and solid tumors, highlighting its potential as a therapeutic agent [196].

WT1-targeted immunotherapies—including monoclonal antibodies, T cell engagers, TCR-mimic antibodies, and bispecific antibodies—demonstrate strong preclinical efficacy and specificity. These advances support their



continued development as targeted therapies for WT1-positive malignancies.

Chimeric antigen receptor (CAR) T-cell therapy targeting WT1

Traditional CAR-T cells target surface antigens; however, WT1 is an intracellular protein. Researchers have engineered CAR-T cells to recognize WT1 peptides presented on MHC class I molecules, overcoming the challenge of WT1's intracellular localization. These engineered T cells demonstrated antitumor activity in xenograft models, with enhanced efficacy when combined with dendritic cell (DC) vaccination, suggesting a synergistic approach to boost CAR-T cell expansion and activation [69, 197].

T-cell Receptor (TCR) gene therapy

Zhang and Li (2019) review WT1-specific TCR gene therapy, where T cells are engineered to express high-affinity TCRs that selectively recognize WT1, a protein overexpressed in cancer cells but limited in normal tissues. This strategy enhances immune targeting and elimination of WT1-positive tumors. Notably, CD4+ T cells engineered with TCRs against the WT1_332 peptide exhibit potent anti-leukemic activity in vitro, highlighting their potential for treating hematologic malignancies [198].

Gielis S et al. analyze the WT1-specific T-cell receptor repertoire in AML patients, revealing greater TCR diversity in remission compared to relapse. These findings highlight the therapeutic potential of diverse WT1-specific TCRs for adoptive T cell therapy. Enhanced TCR diversity may contribute to improved immune control and durable remission in AML [199].

Researchers have engineered high-affinity TCRs that selectively recognize WT1 peptides presented by MHC molecules on tumor cells, enhancing binding affinity and specificity, which in turn improves anti-tumor activity [200, 201]. Preclinical models demonstrate significant tumor regression following WT1-specific TCR-T cell therapy, and early phase clinical trials report prolonged disease-free survival in some patients [201].

Overall, WT1-specific TCR gene therapy represents a promising targeted immunotherapy, with ongoing studies essential to optimize treatment safety, efficacy, and applicability across diverse WT1-expressing malignancies.

crispr-cas9-enhanced TCR gene editing

The integration of CRISPR-Cas9 technology into TCR gene editing has enabled the development of high-avidity,

WT1-specific TCRs with improved antitumor efficacy. Researchers have utilized CRISPR-Cas9 technology to enhance WT1-specific TCR gene editing by integrating TCRs into the TCR α constant (TRAC) locus and disrupting the TCR β constant (TRBC) locus. This approach prevents mispairing with endogenous TCRs, improving T cell specificity and antitumor function. Engineered T cells demonstrate potent cytotoxicity against WT1-expressing tumor cells from AML, ALL, and glioblastoma in vitro and in vivo, while minimizing off-target toxicity. Together, these findings highlight the promise of CRISPR-enhanced WT1 TCR gene therapy as a targeted and effective treatment strategy for WT1-positive malignancies [202].

Gene silencing approaches

WT1 is a promising tumor-associated antigen and therapeutic target in cancers such as leukemia, breast, and lung cancer. Gene silencing of the WT1 (Wilms' Tumor 1) gene has shown considerable potential in cancer treatment. RNA interference (RNAi) strategies effectively reduce WT1 expression, suppressing tumor cell proliferation and inducing apoptosis. WT1-specific small interfering RNA (siRNA) inhibits leukemic cell growth, supporting WT1-targeted RNA-based therapies in oncology [104, 193, 194, 203].

In breast cancer, WT1 silencing reduces tumor growth and metastasis. Aerosol delivery of PEI-RNAi complexes significantly inhibits lung metastases in preclinical models [101]; similarly, respiratory administration of neutral DOPC liposomal-siRNA (L-siRNA) enhances survival and suppresses pulmonary tumor growth [102]. Recent studies demonstrate that combining WT1 gene silencing with chemotherapy overcomes drug resistance and improves treatment efficacy. A comprehensive review highlights the potential of integrating RNAi with conventional therapies to enhance treatment outcomes [203, 204, 205, 206]. Collectively, WT1-targeted gene silencing holds strong potential to improve clinical outcomes in WT1-expressing malignancies.

Table 4 presents a concise yet comprehensive overview of therapeutic strategies targeting WT1, particularly in its oncogenic role. The "Therapy" column identifies each approach by its core design, while the "Mechanism" column describes the biological pathways modulated by the strategy. The "Development" column indicates the current stage of progress, ranging from preclinical research to Phase I/II clinical trials. Where available, the specific treatment agent or molecule is included. Abbreviations used in the table include Ab (antibody), mAbs (monoclonal antibodies), DCs (dendritic cells), and CTLC (cytotoxic T lymphocytes).



Table 4 WT1 Targeted Therapeutic Strategies

Therapy Name	Mechanism	Development	Agent/Example
WT1 Peptide Vaccine [181, 207]	Induces WT1-specific T-cell responses via synthetic peptides	Phase 2 Clinical Trials	Galinpepimut-S
WT1 mRNA Vaccine [208, 209]	Delivers WT1 mRNA to dendritic cells to stimulate anti-tumor immunity	Phase 1/2 Clinical Trials	WT1 mRNA-transfected DCs
WT1 Protein Vaccine [175]	Uses recombinant WT1 protein to elicit immune response	Phase 1 Clinical Trials	Recombinant WT1 protein
WT1 DNA Vaccine [190, 191]	Plasmid DNA encoding WT1 to induce immune response	Preclinical Studies	WT1 DNA plasmid
WT1 Dendritic Cell Vaccine [68]	Dendritic cells pulsed with WT1 peptides or mRNA to activate CTC	Phase 1/2 Clinical Trials	WT1 peptide-pulsed DCs
WT1 TCR-T Cell Therapy [200]	T cells engineered with WT1-specific T-cell receptors	Phase 1 Clinical Trials	WT1-specific TCR-transduced T cells
WT1 CAR-T Cell Therapy [69]	T cells engineered with chimeric anti- gen receptors targeting WT1	Preclinical Studies	WT1/HLA-A*24:02-specific CAR-T cells
WT1 mAbs Therapy [210]	Antibodies targeting WT1 peptide- HLA complexes	Preclinical Studies	ESKM (TCR-mimic antibody)
WT1 siRNA Therapy [102, 211]	Small interfering RNA to silence WT1 gene expression	Preclinical Studies	WT1-targeted siRNA
WT1 shRNA Therapy [212]	Short hairpin RNA for stable WT1 gene silencing	Preclinical Studies	WT1-targeted shRNA
WT1 CRISPR Gene Editing p [202]	CRISPR-Cas9-mediated knockout of WT1 gene	Preclinical Studies	CRISPR-Cas9 targeting WT1
WT1 Bispecific Ab Therapy [196]	Antibodies binding WT1 and T-cell receptors to direct cytotoxicity	Preclinical Studies	WT1-specific bispecific antibodies

Challenges and future directions

Current clinical trials are evaluating the efficacy and safety of WT1-targeted vaccines and other therapeutic strategies across multiple cancer types. These investigations are crucial for determining the clinical applicability of WT1-based treatments in diverse oncological settings.

However, several challenges hinder the development of WT1-targeted therapies. These include the complex and context-dependent biology of WT1, difficulties in drug design, and challenges in clinical translation.

This review highlights the current challenges and complexities associated with WT1-targeted therapies and underscores the need for continued innovation in their clinical application.

WT1 as a context-dependent regulator in cancer

WT1 functions as either a tumor suppressor or an oncogene depending on the cellular and tissue context, complicating the understanding of its role in tumorigenesis and challenging the development of universal WT1-targeted therapies. To avoid unintended oncogenic or tumor-suppressive effects, therapeutic strategies must consider WT1's specific role within each cancer type. As shown in previous studies [67, 213], this complexity requires a nuanced, cancer-specific

approach to therapy design. Continued mechanistic research is essential to unravel the regulatory networks governing WT1's dual roles and to translate this knowledge into safe and effective clinical applications [213].

Isoform diversity and functional complexity

WT1 isoforms exhibit distinct, context-dependent biological functions, with some promoting tumorigenesis and others acting as tumor suppressors. In ovarian epithelial cells, both WT1 (+ KTS) and (- KTS) isoforms enhance proliferation and migration, with the (+ KTS) isoform exerting a stronger effect on migration [214].

Another study investigated the isoform-specific roles of WT1 in acute myeloid leukemia (AML). The authors reported that the (+ KTS) isoform promotes proliferation and inhibits apoptosis, while the (- KTS) isoform suppresses cell growth and induces apoptosis through activation of Caspase 9 [215]. Goel et al. further demonstrated that (- KTS) isoforms are associated with tumor-suppressive functions, including induction of apoptosis via the p53–Bax–Caspase 9 pathway and promotion of differentiation, while (+ KTS) isoforms drive oncogenesis by enhancing proliferation and blocking apoptosis [154].

These findings underscore the functional complexity of WT1 isoforms and their divergent roles across cancer types.



Table 5 WT1 Gene-Related Therapeutic Agents, their mechanism, and status

Therapy Name	Mechanism	Tumors Targeted	Company/Study Name	Status
WT1 Peptide Vaccine (Single Epitope) [216]	Elicits WT1-specific CD8+ cytotoxic T lymphocyte (CTL) response	AML, MDS, NSCLC	Multiple academic consortia	Phase I/II Completed
Galinpepimut-S [170]	Multi-epitope WT1 peptide vaccine stimulating T-cell response	Mesothelioma, Ovarian, AML	SELLAS Life Sciences	Phase II ongoing
OCV-501 [217, 171]	WT1-332 helper peptide vaccine for CD4 ⁺ T-cell activation	AML	OncoTherapy Science	Phase I Completed
WT1-Pulsed Dendritic Cell Vaccine [218, 219]	Dendritic cells presenting WT1 epitopes to activate T cells	Pancreatic, Esophageal, Glioma	National Cancer Center Japan	Pilot/early trials
WT1 CAR T Cells (TCR/ CAR-based) [69]	T cells engineered with WT1-targeting CARs (HLA-restricted)	Leukemia (AML, ALL), Sarcoma	City of Hope/MSKCC	Preclinical/Early phase
Oral WT1 Vaccine (B. longum vector) [220]	Recombinant <i>B. longum</i> displaying WT1 for mucosal immunization	Colorectal (mouse model)	Japanese research consortia	Preclinical (mouse)
WT1 Vaccine + Gemcitabine [188, 221]	Combination of WT1 peptide vaccination with chemo-induced immunomodulation	Pancreatic cancer	Osaka University	Phase I/II Completed
WT1 Vaccine in Pediatric Glioma [222]	WT1 peptide administered in children for glioma/ rhabdomyosarcoma	Pediatric diffuse midline glioma, RMS	Osaka University/Japan Pediatric Study	Case series/feasibility
WT1 CD4 ⁺ T Cell Therapy [170]	HLA-class II-restricted helper T cells targeting WT1	AML	Stanford University	Experimental/Preclinical
WT1 Vaccination Post-allo- HSCT[223]	Enhancing GvL effect via post-transplant WT1 immunotherapy	AML, MDS	Osaka University Hospital	Phase I/II
WT1 Long Peptide Vaccine[176, 181]	Long peptides induce both CD4 ⁺ and CD8 ⁺ responses	Solid tumors, Gliomas	National Cancer Center Japan	Phase I
WT1 TriMix mRNA Vaccine[224, 225]	WT1 mRNA electroporated into DCs+costimulatory signals (CD40L, CD70, TLR4)	AML, MDS	VUB/UZ Brussels	Phase I (early reports)

Selectively targeting oncogenic isoforms while preserving tumor-suppressive variants remains a key therapeutic challenge. Further investigation is required to elucidate the regulatory mechanisms governing WT1 isoform expression and function in cancer (Table 5).

Challenges in drug development

Therapeutic targeting of WT1 remains challenging due to its classification as an "undruggable" transcription factor. Its DNA-binding domain and protein—protein interaction interfaces are poorly accessible to small-molecule inhibitors. Moreover, WT1's nuclear localization complicates intracellular drug delivery. Recent advances in nanotechnology have introduced promising strategies, such as nucleus-targeting WT1 antagonistic peptides encapsulated in polymeric

nanomicelles. While these approaches show potential, they remain in early stages of development [226].

Limited understanding of WT1's role in tumor microenvironment

The role of WT1 in the tumor microenvironment, especially its interactions with immune cells and tumor-associated factors, remains poorly characterized. This knowledge gap hampers the development of targeted therapies that effectively modulate the tumor microenvironment to improve outcomes. Immunosuppression within the tumor microenvironment significantly limits WT1 vaccine efficacy by suppressing antitumor immune responses. To enhance vaccine performance, researchers are investigating adjuvants, combination therapies, and genetic modifications that enhance antigen



presentation. Recent studies also highlight nanoparticle-based vaccines as a promising approach to overcome tumor-induced immunosuppression and improve therapeutic success in solid tumors [227].

Heterogeneity in WT1 expression across tumor types

Significant intra- and inter-tumoral heterogeneity in WT1 expression limits its utility as a universal biomarker and therapeutic target, complicating standardized WT1-targeted treatment development, particularly in tumors with low or transient WT1 levels. Van Amerongen et al. identified novel immunogenic WT1-derived peptides and engineered highaffinity T-cell receptors (TCRs) that enhance antitumor responses in acute myeloid leukemia and ovarian carcinoma [128]. Recent studies focus on enhancing the delivery and precision of WT1-targeted therapies. Researchers apply RNA interference (RNAi) and small interfering RNA (siRNA) to suppress WT1 expression, thereby inhibiting tumor growth and improving combination therapy efficacy. Additionally, CRISPR-based gene editing and WT1 peptide vaccines are under investigation to elicit targeted immune responses against WT1-expressing tumor cells. These approaches aim to overcome WT1 expression heterogeneity and enable more effective, personalized WT1-directed immunotherapies [103].

Immunogenicity and safety concerns

WT1-targeted vaccines have demonstrated safety and immunogenicity; however, eliciting durable and robust immune responses remains a challenge. In a first-in-human study, Kreutmair et al. (2022) administered WT1 recombinant protein vaccines to elderly AML patients in remission. Immune responses varied: Some patients maintained durable remissions, while others relapsed, often with clonal evolution, indicating the need for improved vaccine strategies [175].

Although early phase trials rarely report autoimmune events, emerging data suggest immune-related risks under certain conditions. Suzuki et al. (2019) documented newonset autoimmune disorders in patients with refractory thymic epithelial malignancies receiving WT1 peptide immunotherapy. These findings highlight the necessity for stringent patient selection and close monitoring during WT1-targeted treatment [228].

The spontaneous clonal expansion of WT1-specific cytotoxic T lymphocytes in patients with WT1-expressing tumors highlights the strong immunogenicity of the WT1 antigen, with no evidence of systemic autoimmunity, supporting its potential as a safe and effective target for immunotherapy [229].

Inhibition of the immunoproteasome with ONX-0914 enhances processing and presentation of the WT1_235 epitope in mesothelioma cells, leading to increased recognition by WT1-specific cytotoxic T lymphocytes and thereby boosting tumor immunogenicity [230].

Variability in clinical responses and challenges in monitoring treatment efficacy

Clinical trials investigating WT1-targeted therapies have shown mixed results. For example, Suzuki et al. (2019) reported that only 46.2% of patients with refractory thymic epithelial malignancies developed WT1-specific IgG antibodies after peptide-based immunotherapy, with no significant tumor shrinkage observed. These findings highlight the need for more potent and broadly effective WT1-targeted approaches [228].

To address such limitations, Lahman et al. (2022) designed an alternative WT1 peptide that bypasses immunoproteasome dependency, aiming to enhance antigen presentation and potentially improve immune responses [231].

Monitoring WT1 expression levels to assess treatment response is complicated by variability across malignancies and patient populations. Deng et al. (2021) highlighted the challenges in monitoring WT1 expression levels for assessing treatment response in pediatric acute myeloid leukemia (AML), noting that WT1 expression can fluctuate, complicating its use as a reliable biomarker for minimal residual disease detection or relapse prediction [232].

These findings underscore the variability in clinical responses to WT1-targeted therapies and the challenges in monitoring treatment efficacy, emphasizing the need for improved strategies and biomarkers.

On-target, off-tumor toxicity

WT1-targeted immunotherapies pose a risk of on-target, off-tumor toxicity due to WT1's low-level expression in normal tissues such as the kidneys and gonads. WT1-specific CAR-T and TCR-engineered T cells can recognize and damage both malignant and healthy WT1-expressing cells, leading to unintended cytotoxicity. Clinical studies have documented such risks in patients with AML and MDS treated with WT1-specific TCR-transduced lymphocytes. Akahori et al. further demonstrated that WT1-targeted CAR-T cells can exhibit potent antitumor effects, but emphasized the need for enhanced specificity to mitigate off-tumor toxicity. These findings underscore the need for refined targeting strategies that maintain antitumor efficacy while minimizing collateral damage to normal WT1-expressing cells [69, 200].



336

Challenges in biomarker development

Reliable biomarkers for monitoring WT1-targeted therapies in Wilms tumor remain insufficiently developed. Liquid biopsy methods, including circulating tumor DNA and exosomes, demonstrate potential for early detection and disease monitoring but face limited clinical implementation. Existing studies report inconsistent findings and lack of comprehensive validation of their sensitivity and specificity. This highlights the urgent need for comprehensive research and rigorous clinical validation to establish their accuracy. Advancing biomarker development is critical to enable personalized treatment and improve outcomes in Wilms tumor patients. Despite their promise, liquid biopsies require further validation before routine clinical use [169].

Inadequate clinical trial infrastructure

Preclinical studies, case reports, and early phase trials of WT1-targeted therapies have demonstrated safety and potential efficacy; however, large-scale multicenter clinical trials remain lacking. This evidentiary gap limits the clinical integration of WT1-targeted treatments.

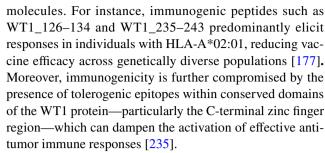
Most existing studies are retrospective with small cohorts, restricting result generalizability. For instance, Boublikova et al. (2006) observed substantial variability in WT1 expression in childhood acute lymphoblastic leukemia (ALL), but the limited sample size prevented definitive prognostic conclusions. This highlights the need for larger, well-designed trials to better understand WT1's clinical significance [233].

A recent large-scale analysis of 533 pediatric B-cell precursors ALL (BCP-ALL) patients reported a broad range of WT1 transcript levels (median 0.26%) and identified WT1 overexpression with poorer overall and event-free survival. However, WT1 overexpression did not independently predict outcomes across the entire cohort, highlighting the complexity and variability in WT1's prognostic significance across different patient subgroups [234].

These findings emphasize the necessity for large-scale, multicenter clinical trials to clarify WT1's prognostic role in childhood ALL and to standardize its clinical application. Strengthening clinical trial infrastructure is essential to validate WT1's significance and guide therapeutic development.

HLA Restriction in WT1-peptide-based vaccines

WT1 peptide-based vaccines exhibit immunogenicity; however, their clinical applicability is constrained across diverse populations by interindividual variability in human leukocyte antigen (HLA) expression. This variability can influence the effectiveness of WT1 peptide vaccines, as the immunogenicity of these vaccines depends on the patient's ability to present the WT1-derived peptides via their HLA



To overcome HLA restriction and tolerance, novel strategies are being explored. For instance, next-generation DNA vaccine platforms encoding truncated or modified WT1 sequences. These constructs enhance epitope presentation, break immune tolerance, and induce neoantigen-specific T cell responses, thereby broadening patient eligibility independent of HLA subtype [177, 190, 191, 235, 236]. These findings underscore the need to optimize WT1-based vaccines by integrating strategies that expand HLA coverage and overcome intrinsic immune tolerance mechanisms.

Advancements in genomics and personalized medicine offer the opportunity to develop individualized WT1 treatment strategies of each patient. Further research is needed to elucidate the molecular mechanisms underlying the dual functions of WT1 and to identify biomarkers that can predict patient response to WT1-targeted therapies. A deeper understanding of these mechanisms will facilitate the development of more effective and personalized cancer treatment strategies.

Conclusion

The Wilms' Tumor 1 (WT1) gene plays a pivotal role in cancer biology, acting as both a tumor suppressor and oncogene depending on cellular context. Its regulation of key molecular pathways, interaction with tumor suppressors, and impact on gene expression highlight its significance in cancer pathogenesis. Aberrant WT1 expression in malignancies such as leukemia, lung, and ovarian cancers establishes it as a prognostic marker and a promising therapeutic target. WT1-directed strategies, including vaccines and molecular therapies, show potential to enhance treatment efficacy. Advances in immunotherapy and molecular research continue to refine these approaches. Ongoing clinical trials and focused research are essential to overcoming current challenges and maximizing the therapeutic value of WT1 in cancer management.

Acknowledgements We would like to thank the reviewers and editors for their valuable feedback and guidance in improving the manuscript.

Author contributions Authors Contributions: R.Q1. Designed the study, conducted the research, and wrote the manuscript. T.H.



supervised project administration. A.Q. contributed to the literature review and creation of illustrations. S.Q. and R.Q2 offered valuable insights into the data presentation. All authors have read and approved the final version of the manuscript.

Funding The author(s) received no specific grant from any public, commercial, or not-for-profit agencies for the research, authorship, or publication of this article.

Data availability No datasets were generated or analysed during the current study.

Declarations

Conflict of interest The authors declare no competing interests.

Ethics approval and consent to participate This manuscript does not involve the use of animal or human data or tissue; therefore, ethical approval is not required in this section.

Open Access This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by-nc-nd/4.0/.

References

- Bray F, Laversanne M, Weiderpass E, Soerjomataram I. The everincreasing importance of cancer as a leading cause of premature death worldwide. Cancer. 2021;127(16):3029–30. https://doi.org/ 10.1002/cncr.33587.
- Bray F, Laversanne M, Sung H, Ferlay J, Siegel RL, Soerjomataram I, et al. Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2024;74(3):229–63. https://doi.org/10.3322/caac.21834.
- Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011;144(5):646–74. https://doi.org/10.1016/j.cell. 2011.02.013.
- Estanqueiro M, Amaral MH, Conceição J, Sousa Lobo JM. Nanotechnological carriers for cancer chemotherapy: the state of the art. Colloids Surf B Biointerfaces. 2015;126:631–48. https://doi. org/10.1016/j.colsurfb.2014.12.041.
- Diori Karidio I, Sanlier SH. Reviewing cancer's biology: an eclectic approach. J Egypt Natl Canc Inst. 2021;33(1):32. https:// doi.org/10.1186/s43046-021-00088-y.
- Loftus LV, Amend SR, Pienta KJ. Interplay between cell death and cell proliferation reveals new strategies for cancer therapy. Int J Mol Sci. 2022;23(9):4723. https://doi.org/10.3390/ijms2 3094723.

- Quraish RU, Hirahata T, Quraish AU, Ul Quraish S. An overview: genetic tumor markers for early detection and current gene therapy strategies. Cancer Inform. 2023;22:11769351221150772. https://doi.org/10.1177/11769351221150772.
- Essafi A, Hastie ND. WT1 the oncogene: a tale of death and HtrA. Mol Cell. 2010;37(2):153–5. https://doi.org/10.1016/j. molcel.2010.01.010.
- Kontomanolis EN, Koutras A, Syllaios A, Schizas D, Mastoraki A, Garmpis N, et al. Role of oncogenes and tumorsuppressor genes in carcinogenesis: a review. Anticancer Res. 2020;40(11):6009–15. https://doi.org/10.21873/anticanres. 14622.
- Dakal TC, Dhabhai B, Pant A, Moar K, Chaudhary K, Yadav V, et al. Oncogenes and tumor suppressor genes: functions and roles in cancers. MedComm. 2024;5(6):e582. https://doi.org/10.1002/ mco2.582.
- Haber DA, Buckler AJ, Glaser T, Call KM, Pelletier J, Sohn RL, et al. An internal deletion within an 11p13 zinc finger gene contributes to the development of Wilms' tumor. Cell. 1990;61(7):1257–69. https://doi.org/10.1016/0092-8674(90) 90690-g.
- Treger TD, Chowdhury T, Pritchard-Jones K, Behjati S. The genetic changes of Wilms tumour. Nat Rev Nephrol. 2019;15(4):240–51. https://doi.org/10.1038/s41581-019-0112-0.
- 13. Ruteshouser EC, Robinson SM, Huff V. Wilms tumor genetics: mutations in WT1, WTX, and CTNNB1 account for only about one-third of tumors. Genes Chromosomes Cancer. 2008;47(6):461–70. https://doi.org/10.1002/gcc.20553.
- 14. Anitha GRJ, Chandraprabha VR, Padmakumar D, Sreelatha MM, Padmakumar A, Gopinath P, et al. Exploring the prevalence and prognostic impact of Wilms tumor 1 exon 7 mutation status with combinations of FLT3-ITD and NPM1 mutations as potential molecular biomarkers in acute myeloid leukemia patients with normal cytogenetics. Asian Pac J Cancer Prev. 2024;25(10):3463–70. https://doi.org/10.31557/APJCP.2024. 25.10.3463.
- 15. Huff V. Wilms' tumours: about tumour suppressor genes, an oncogene and a chameleon gene. Nat Rev Cancer. 2011;11(2):111–21. https://doi.org/10.1038/nrc3002.
- Jing YJ, Lin LC, Chen LL, et al. WT1 inhibits human renal carcinoma cell proliferation and induces G2/M arrest by upregulating IL-24 expression. Biomed Res Int. 2022;2022:1093945. https://doi.org/10.1155/2022/1093945.
- Rampal R, Figueroa ME. Wilms tumor 1 mutations in the pathogenesis of acute myeloid leukemia. Haematologica. 2016;101(6):672-9. https://doi.org/10.3324/haematol.2015. 141796
- McGregor RJ, Chau Y, Kendall TJ, Artibani M, Hastie N, Hadoke PWF. WT1 expression in vessels varies with histopathological grade in tumour-bearing and control tissue from patients with breast cancer. Br J Cancer. 2018;119:1508–17. https://doi.org/ 10.1038/s41416-018-0317-1.
- 19. Liu Z, Yamanouchi K, Ohtao T, et al. High levels of Wilms' tumor 1 (WT1) expression were associated with aggressive clinical features in ovarian cancer. Anticancer Res. 2014;34(5):2331–40.
- Novacescu D, Cut TG, Cumpanas AA, Latcu CS, Bardan R, Ferician O, et al. Evaluating established roles, future perspectives and methodological heterogeneity for Wilms' Tumor 1 (WT1) antigen detection in adult renal cell carcinoma, using a novel N-terminus targeted antibody (clone WT49). Biomedicines. 2022;10(4):912. https://doi.org/10.3390/biomedicines10040912.
- Zapata-Benavides P, Manilla-Muñoz E, Zamora-Avila DE, et al. WT1 silencing by RNAi synergizes with chemotherapeutic agents and induces chemosensitization to doxorubicin



- and cisplatin in B16F10 murine melanoma cells. Oncol Lett. 2012;3(4):751–5. https://doi.org/10.3892/ol.2012.578.
- Kijima N, Hosen N, Kagawa N, Kinoshita M, Oji Y, et al. Wilms' tumor 1 is involved in tumorigenicity of glioblastoma by regulating cell proliferation and apoptosis. Anticancer Res. 2014;34(1):61-7.
- Kijima N, Hashimoto N, Chiba Y, Fujimoto Y, Sugiyama H, Yoshimine T. Functional roles of Wilms' Tumor 1 (WT1) in malignant brain tumors. In: van den Heuvel-Eibrink MM, ed. Wilms Tumor. Brisbane (AU): Codon Publications; 2016. Chapter 15. https://doi.org/10.15586/codon.wt.2016.ch15.
- Henon C, Vibert J, Eychenne T, Colmet-Daage L, Ngo C, Garrido M, et al. Single-cell multiomics profiling reveals heterogeneous transcriptional programs and microenvironment in DSRCTs. Cell Rep Med. 2024;5(6):101582. https://doi.org/10.1016/j.xcrm. 2024 101582.
- Sugiyama H. WT1 (Wilms' Tumor Gene 1): biology and Cancer Immunotherapy. Jpn J Clin Oncol. 2010;40(5):377–87. https://doi.org/10.1093/jjco/hyp194.
- Dong L, Pietsch S, Englert C. Towards an understanding of kidney diseases associated with WT1 mutations. Kidney Int. 2015;88(4):684–90. https://doi.org/10.1038/ki.2015.198.
- 27. Stoll R, Lee BM, Debler EW, Laity JH, Wilson IA, Dyson HJ, et al. Structure of the Wilms tumor suppressor protein zinc finger domain bound to DNA. J Mol Biol. 2007;372(5):1227–45.
- 28. Ullmark T, Järvstråt L, Sandén C, Montano G, Jernmark-Nilsson H, Lilljebjörn H, et al. Distinct global binding patterns of the Wilms tumor gene 1 (WT1) -KTS and +KTS isoforms in leukemic cells. Haematologica. 2017;102(2):336–45. https://doi.org/10.3324/haematol.2016.149815.
- Lee KY, Jeon YJ, Kim HG, Ryu J, Lim DY, Jung SK, et al. The CUG-translated WT1, not AUG-WT1, is an oncogene. Carcinogenesis. 2017. https://doi.org/10.1093/carcin/bgx108.
- 30. Ozdemir DD, Hohenstein P. WT1 in the kidney—a tale in mouse models. Pediatr Nephrol. 2014;29:687–93. https://doi.org/10.1007/s00467-013-2673-7.
- 31. Benetti E, Caridi G, Malaventura C, Dagnino M, Leonardi E, Artifoni L, et al. A novel WT1 gene mutation in a three-generation family with progressive isolated focal segmental glomerulosclerosis. Clin J Am Soc Nephrol. 2010;5(4):698–702. https://doi.org/10.2215/CJN.05670809.
- 32. Ullmark T, Montano G, Gullberg U. DNA and RNA binding by the Wilms' tumour gene 1 (WT1) protein +KTS and -KTS isoforms—from initial observations to recent global genomic analyses. Eur J Haematol. 2018;100(3):229–40. https://doi.org/10.1111/ejh.13010.
- Wells J, Rivera MN, Kim WJ, Starbuck K, Haber DA. The predominant WT1 isoform (+KTS) encodes a DNA-binding protein targeting the planar cell polarity gene Scribble in renal podocytes. Mol Cancer Res. 2010;8(7):975–85. https://doi.org/ 10.1158/1541-7786.MCR-10-0033.
- 34. Wang L, Zhang X, Wang ZY. The Wilms' tumor suppressor WT1 regulates expression of members of the epidermal growth factor receptor (EGFR) and estrogen receptor in acquired tamoxifen resistance. Anticancer Res. 2010;30(9):3637-42.
- 35. Yang C, Romaniuk PJ. The ratio of +/-KTS splice variants of the Wilms' tumour suppressor protein WT1 mRNA is determined by an intronic enhancer. Biochem Cell Biol. 2008;86(4):312-21. https://doi.org/10.1139/o08-075.
- Parenti R, Puzzo L, Vecchio GM, Gravina L, Salvatorelli L, Musumeci G, et al. Immunolocalization of Wilms' tumor protein (WT1) in developing human peripheral sympathetic and gastroenteric nervous system. Acta Histochem. 2014;116(1):48-54. https://doi.org/10.1016/j.acthis.2013.05.003.

- Parenti R, Perris R, Vecchio GM, Salvatorelli L, Torrisi A, Gravina L, et al. Immunohistochemical expression of Wilms' tumor protein (WT1) in developing human epithelial and mesenchymal tissues. Acta Histochem. 2013;115(1):70–5. https:// doi.org/10.1016/j.acthis.2012.04.006.
- Parenti R, Salvatorelli L, Musumeci G, Parenti C, Giorlandino A, Motta F, et al. Wilms' tumor 1 (WT1) protein expression in human developing tissues. Acta Histochem. 2015;117(4–5):386–96. https://doi.org/10.1016/j.acthis.2015.03.009.
- Miller-Hodges E, Hohenstein P. WT1 in disease: shifting the epithelial-mesenchymal balance. J Pathol. 2012;226(2):229– 40. https://doi.org/10.1002/path.2977.
- Kim MS, Hancock AL, Harper JC, Park JY, Poy G, Perantoni AO, et al. Identification of novel Wilms' tumor suppressor gene target genes implicated in kidney development. J Biol Chem. 2007;282(22):16278–87. https://doi.org/10.1074/jbc.M7002 15200.
- Kim MK, McGarry TJ, O Broin P, Flatow JM, Golden AA, Licht JD. An integrated genome screen identifies the Wnt signaling pathway as a major target of WT1. Proc Natl Acad Sci USA. 2009;106(27):11154–9. https://doi.org/10.1073/pnas. 0901591106.
- Roberts SGE. Transcriptional regulation by WT1 in development. Curr Opin Genet Dev. 2005;15(5):542–7. https://doi.org/10.1016/j.gde.2005.08.004.
- Essafi A, Webb A, Berry RL, Slight J, Burn SF, Spraggon L, et al. A WT1-controlled chromatin switching mechanism underpins tissue-specific Wnt4 activation and repression. Dev Cell. 2011;21(3):559–74. https://doi.org/10.1016/j.devcel.2011.07. 014
- 44. Toska E, Roberts SGE. Mechanisms of transcriptional regulation by WT1 (Wilms' tumour 1). Biochem J. 2014;461(1):15–32. https://doi.org/10.1042/BJ20131587.
- McKay LM, Carpenter B, Roberts SGE. Regulation of the Wilms' tumour suppressor protein transcriptional activation domain. Oncogene. 1999;18(47):6546–54. https://doi.org/10. 1038/sj.onc.1203046.
- Scharnhorst V, van der Eb AJ, Jochemsen AG. WT1 proteins: functions in growth and differentiation. Gene. 2001;273(2):141– 61. https://doi.org/10.1016/S0378-1119(01)00593-5.
- Lee TH, Moffett P, Pelletier J. The Wilms' tumor suppressor gene (WT1) product represses different functional classes of transcriptional activation domains. Nucleic Acids Res. 1999;27(14):2889–97. https://doi.org/10.1093/nar/27.14.2889.
- Carpenter B, Hill KJ, Charalambous M, Wagner KJ, Lahiri D, James DI, et al. BASP1 is a transcriptional cosuppressor for the Wilms' tumor suppressor protein WT1. Mol Cell Biol. 2004;24(2):537–49. https://doi.org/10.1128/MCB.24.2.537-549.
- Sugiyama H. Wilms' tumor gene WT1: its oncogenic function and clinical application. Int J Hematol. 2001;73(2):177–87. https://doi.org/10.1007/BF02981935.
- Ferrari MT, Elias FM, Gomes NL, Batista RL, Faria JA Jr, Nishi MY. WT1: A single gene associated with multiple and severe phenotypes. Endocrine and Metabolic Science. 2023;13:100143. https://doi.org/10.1016/j.endmts.2023.100143.
- Hastie ND. Wilms' tumour 1 (WT1) in development, homeostasis and disease. Development. 2017;144(16):2862–72. https://doi.org/10.1242/dev.153163.
- Del Díaz Moral S, Wagner N, Wagner KD. The Wilms' tumor suppressor WT1 in cardiomyocytes: implications for cardiac homeostasis and repair. Cells. 2024;13(24):2078. https://doi. org/10.3390/cells13242078.
- Quenneville J, Feghaly A, Tual M, et al. Long-term severe hypoxia adaptation induces non-canonical EMT and a



- novel Wilms tumor 1 (WT1) isoform. Cancer Gene Ther. 2024;31:1237–50. https://doi.org/10.1038/s41417-024-00795-3.
- Waehle V, Ungricht R, Hoppe PS, Betschinger J. The tumor suppressor WT1 drives progenitor cell progression and epithelialization to prevent Wilms tumorigenesis in human kidney organoids. Stem Cell Reports. 2021;16(9):2107–17. https://doi.org/10.1016/j.stemcr.2021.07.023.
- Larsson SH, Charlieu JP, Miyagawa K, Engelkamp D, Rassoulzadegan M, Ross A, et al. Subnuclear localization of WT1 in splicing or transcription factor domains is regulated by alternative splicing. Cell. 1995;81(3):391–401. https://doi.org/10.1016/0092-8674(95)90392-5.
- Dudnakova T, Spraggon L, Slight J, Hastie N. Actin: a novel interaction partner of WT1 influencing its cell dynamic properties. Oncogene. 2010;29(7):1085–92. https://doi.org/10.1038/ onc.2009.444.
- 57. Niksic M, Slight J, Sanford J, Caceres JF, Hastie ND. The Wilms' tumour protein (WT1) shuttles between nucleus and cytoplasm and is present in functional polysomes. Hum Mol Genet. 2004;13(4):463–71. https://doi.org/10.1093/hmg/ddh040.
- Wegert J, Ishaque N, Vardapour R, Geörg C, Gu Z, Bieg M, et al. Mutations in the SIX1/2 pathway and the DROSHA/DGCR8 miRNA microprocessor complex underlie high-risk blastemal type Wilms tumors. Cancer Cell. 2015;27(2):298–311. https:// doi.org/10.1016/j.ccell.2015.01.002.
- Ben Haj Othmen H, Othman H, Khamessi O, Bettaieb I, Gara S, Kharrat M. Overexpression of WT1 in all molecular subtypes of breast cancer and its impact on survival: exploring oncogenic and tumor suppressor roles of distinct WT1 isoforms. Mol Biol Rep. 2024;51(1):544. https://doi.org/10.1007/s11033-024-09450-4.
- Hohenstein P, Hastie ND. The many facets of the Wilms' tumour gene, WT1. Hum Mol Genet. 2006;15(Spec No 2):R196–201. https://doi.org/10.1093/hmg/ddl196.
- Chau YY, Hastie ND. The role of WT1 in regulating mesenchyme in cancer, development, and tissue homeostasis. Trends Genet. 2012;28(10):515–24. https://doi.org/10.1016/j.tig.2012.04.004.
- Yao Y, Chai X, Gong C, Zou L. WT1 inhibits AML cell proliferation in a p53-dependent manner. Cell Cycle. 2021;20(16):1552–60. https://doi.org/10.1080/15384101.2021.1951938.
- Yang L, Han Y, Suarez Saiz F, Minden MD. A tumor suppressor and oncogene: the WT1 story. Leukemia. 2007;21(5):868–76. https://doi.org/10.1038/sj.leu.2404624.
- 64. Ito K, Oji Y, Tatsumi N, Shimizu S, Kanai Y, Nakazawa T, Asada M, et al. Antiapoptotic function of 17AA(+)WT1 (Wilms' tumor gene) isoforms on the intrinsic apoptosis pathway. Oncogene. 2006;25(30):4217–29. https://doi.org/10.1038/sj.onc.1209455.
- 65. Bansal H, Seifert T, Bachier C, Rao M, Tomlinson G, Iyer SP, et al. The transcription factor Wilms tumor 1 confers resistance in myeloid leukemia cells against the proapoptotic therapeutic agent TRAIL by regulating the antiapoptotic protein Bcl-xL. J Biol Chem. 2012;287(39):32875–80. https://doi.org/10.1074/jbc. C112.366559.
- Simpson LA, Burwell EA, Thompson KA, Shahnaz S, Chen AR, Loeb DM. The antiapoptotic gene A1/BFL1 is a WT1 target gene that mediates granulocytic differentiation and resistance to chemotherapy. Blood. 2006;107(12):4695–702. https://doi.org/ 10.1182/blood-2005-10-4025.
- Qing N, Lin Y, Zeng J, Zhang Y, Liu R. Multifaceted functions of the Wilms tumor 1 protein: from its expression in various malignancies to targeted therapy. Transl Oncol. 2025;52:102237. https://doi.org/10.1016/j.tranon.2024.102237.
- Berneman ZN, De Laere M, Germonpré P, et al. WT1-mRNA dendritic cell vaccination of patients with glioblastoma multiforme, malignant pleural mesothelioma, metastatic breast cancer,

- and other solid tumors: type 1 T-lymphocyte responses are associated with clinical outcome. J Hematol Oncol. 2025;18(1):9. https://doi.org/10.1186/s13045-025-01661-x.
- Akahori Y, Wang L, Yoneyama M, et al. Antitumor activity of CAR-T cells targeting the intracellular oncoprotein WT1 can be enhanced by vaccination. Blood. 2018;132(11):1134

 45. https://doi.org/10.1182/blood-2017-08-802926.
- 70. Maheswaran S, Englert C, Zheng G, et al. Inhibition of cellular proliferation by the Wilms tumor suppressor WT1 requires association with the inducible chaperone Hsp70. Genes Dev. 1998;12(8):1108–20. https://doi.org/10.1101/gad.12.8.1108.
- 71. Maheswaran S, Englert C, Bennett P, Heinrich G, Haber DA. The WT1 gene product stabilizes p53 and inhibits p53-mediated apoptosis. Genes Dev. 1995;9(17):2143–56. https://doi.org/10.1101/gad.9.17.2143.
- Bordin F, Piovan E, Masiero E, et al. WT1 loss attenuates the TP53-induced DNA damage response in T-cell acute lymphoblastic leukemia. Haematologica. 2018;103(2):266–77. https:// doi.org/10.3324/haematol.2017.170431.
- Stanhope-Baker P, Kessler PM, Li W, Agarwal ML, Williams BR. The Wilms tumor suppressor-1 target gene podocalyxin is transcriptionally repressed by p53. J Biol Chem. 2004;279(32):33575–85. https://doi.org/10.1074/jbc.M404787200.
- 74. Inoue K, Fry EA. Alterations of p63 and p73 in human cancers. Subcell Biochem. 2014;85:17–40. https://doi.org/10.1007/978-94017-9211-0_2.
- 75. Scharnhorst V, Dekker P, van der Eb AJ, Jochemsen AG. Physical interaction between Wilms tumor 1 and p73 proteins modulates their functions. J Biol Chem. 2000;275(14):10202–11. https://doi.org/10.1074/jbc.275.14.10202.
- Li X, Ottosson S, Wang S, et al. Wilms' tumor gene 1 regulates p63 and promotes cell proliferation in squamous cell carcinoma of the head and neck. BMC Cancer. 2015;15(1):342. https://doi. org/10.1186/s12885-015-1356-0.
- Wu C, Yang J, Li R, Lin X, Wu J, Wu J. LncRNA WT1-AS/miR-494–3p regulates cell proliferation, apoptosis, migration and invasion via PTEN/PI3K/AKT signaling pathway in non-small cell lung cancer. *Onco Targets Ther.* 2021;14: 891–904. Published 2021 Feb 9. [Retracted in: *Onco Targets Ther.* 2024; 17:739–740. https://doi.org/10.2147/OTT.S278233.
- Chen W, Zhuang J, Gong L, Dai Y, Diao H. Investigating the dysfunctional pathogenesis of Wilms' tumor through a multidimensional integration strategy. Ann Transl Med. 2019;7(7):136. https://doi.org/10.21037/atm.2019.03.37.
- Liu GL, Yang HJ, Liu B, Liu T. Effects of microRNA-19b on the proliferation, apoptosis, and migration of Wilms' tumor cells via the PTEN/PI3K/AKT signaling pathway. *J Cell Biochem*. 2017;118(10):3424–3434. [Retracted in: *J Cell Biochem*. 2021 Nov;122(Suppl 1):S55. https://doi.org/10.1002/jcb.25999.
- Witus SR, Tuttle LM, Li W, et al. BRCA1/BARD1 intrinsically disordered regions facilitate chromatin recruitment and ubiquitylation. EMBO J. 2023;42(15):e113565. https://doi.org/10.15252/ embj.2023113565.
- Wegert J, Appenzeller S, Treger TD, et al. Distinct pathways for genetic and epigenetic predisposition in familial and bilateral Wilms tumor. Genome Med. 2025;17(1):49. https://doi.org/10. 1186/s13073-025-01482-0.
- Rather MI, Swamy S, Gopinath KS, Kumar A. Transcriptional repression of tumor suppressor CDC73, encoding an RNA polymerase II interactor, by Wilms tumor 1 protein (WT1) promotes cell proliferation: implication for cancer therapeutics. J Biol Chem. 2014;289(2):968–76. https://doi.org/10.1074/jbc.M113. 483255.



- Mayo MW, Wang CY, Drouin SS, et al. WT1 modulates apoptosis by transcriptionally upregulating the Bcl-2 protooncogene. EMBO J. 1999;18(14):3990–4003. https://doi.org/10.1093/emboj/18.14.3990.
- 84. Morrison DJ, English MA, Licht JD. WT1 induces apoptosis through transcriptional regulation of the proapoptotic Bcl2 family member Bak. Cancer Res. 2005;65(18):8174–82. https://doi.org/10.1158/0008-5472.CAN-04-3657.
- Cheema SK, Mishra SK, Rangnekar VM, Tari AM, Kumar R, Lopez-Berestein G. Par-4 transcriptionally regulates Bcl-2 through a WT1-binding site on the Bcl-2 promoter. J Biol Chem. 2003;278(22):19995–20005. https://doi.org/10.1074/jbc.M205865200.
- Morrison DJ, Kim MK, Berkofsky-Fessler W, Licht JD. WT1 induction of mitogen-activated protein kinase phosphatase 3 represents a novel mechanism of growth suppression. Mol Cancer Res. 2008;6(7):1225–31. https://doi.org/10.1158/15417786. MCR-08-0078.
- 87. Zhang TF, Yu SQ, Guan LS, Wang ZY. Inhibition of breast cancer cell growth by the Wilms' tumor suppressor WT1 is associated with a destabilization of beta-catenin. Anticancer Res. 2003;23:3575–84.
- Dao T, Yan S, Veomett N, et al. Targeting the intracellular WT1 oncogene product with a therapeutic human antibody. Sci Transl Med. 2013;5(176):176ra33. https://doi.org/10.1126/scitr anslmed.3005661.
- 89. Manthopoulou E, Ramai D, Dhar J, et al. Cholangiocarcinoma in the era of immunotherapy. Vaccines (Basel). 2023;11(6):1062.
- Green BL. A peptide vaccine targeting WT1 for cholangiocarcinoma. *Cholangiocarcinoma Foundation 2024 Annual Conference*. Presented April 17–19, 2024. Accessed 6 June 2024.
- Li X, An W, Pan H, et al. Wilms' tumour gene 1 (WT1) enhances non-small cell lung cancer malignancy and is inhibited by micro-RNA-498-5p. BMC Cancer. 2023;23(1):909. https://doi.org/10. 1186/s12885-023-11399-9.
- Nishida S, Morimoto S, Oji Y, et al. Cellular and humoral immune responses induced by an HLA class I-restricted peptide cancer vaccine targeting WT1 are associated with favorable clinical outcomes in advanced ovarian cancer. J Immunother. 2022;45(1):56–66. https://doi.org/10.1097/CJI.0000000000 000405.
- 93. Liu H, Zha Y, Malnassy G, Fulton N. WT1 peptide vaccine is able to induce WT1-specific immune response with TCR clonal enrichment to control minimal residual disease in patients with myeloid leukemia. Blood. 2016;128(22):3984. https://doi.org/10.1182/blood.V128.22.3984.3984.
- Liu H, Zha Y, Choudhury N, et al. WT1 peptide vaccine in Montanide, in contrast to poly ICLC, is able to induce WT1specific immune response with TCR clonal enrichment in myeloid leukemia. Exp Hematol Oncol. 2018;7:1. https://doi.org/10.1186/s40164-018-0093x.
- Nagano C, Nozu K. A review of the genetic background in complicated WT1-related disorders. Clin Exp Nephrol. 2025;29(1):1–9. https://doi.org/10.1007/s10157-024-02539-x.
- Nagano C, Takaoka Y, Kamei K, et al. Genotype-phenotype correlation in WT1 exon 8 to 9 missense variants. Kidney Int Rep. 2021;6(8):2114–21. https://doi.org/10.1016/j.ekir.2021.05.009.
- Lipska BS, et al. Genotype–phenotype associations in WT1 glomerulopathy. Kidney Int. 2014;85(5):1169–78.
- Chen H, Zhang M, Lin J, et al. Genotype-phenotype correlation of WT1 mutation-related nephropathy in Chinese children. Front Pediatr. 2023;11:1192021. https://doi.org/10.3389/fped. 2023.1192021.
- Goel H, Pandey AK, Kumar R, Kumar R, Ningombam SS. RNA sequencing identifies WT1 overexpression as a predictor of poor

- outcomes in acute myeloid leukemia. Cancers. 2025;17(11):1818. https://doi.org/10.3390/cancers17111818.
- 100. Klein C, Augsberger C, Xu W, Heitmüller C, Hanisch L, Sam J, et al. Targeting intracellular WT1 in AML utilizing a T cell bispecific antibody construct: augmenting efficacy through combination with lenalidomide. Blood. 2019;134(Supplement_1):4450-4450. https://doi.org/10.1182/blood-2019-130121.
- Zamora-Avila DE, Zapata-Benavides P, Franco-Molina MA, et al. WT1 gene silencing by aerosol delivery of PEI-RNAi complexes inhibits B16–F10 lung metastases growth. Cancer Gene Ther. 2009;16(12):892–9. https://doi.org/10.1038/cgt.2009.35.
- 102. Ramos-Gonzalez MR, Vazquez-Garza E, Garcia-Rivas G, Rodriguez-Aguayo C, Chavez-Reyes A. Therapeutic effects of WT1 silencing via respiratory administration of neutral DOPC liposomal-siRNA in a lung metastasis melanoma murine model. Noncoding RNA. 2023;9(2):21. https://doi.org/10.3390/ncrna 9020021
- Hein KZ, Yao S, Fu S. Wilms' tumor 1 (WT1): the vaccine for cancer. J Immunother Precis Oncol. 2020;3(4):165–71. https:// doi.org/10.36401/JIPO-20-12.
- Sugiyama H. WT1 cancer vaccine for the treatment and prevention. Med Res Arch. 2022. https://doi.org/10.18103/mra.v10i4. 2762.
- Udtha M, Lee SJ, Alam R, Coombes K, Huff V. Upregulation of c-MYC in WT1-mutant tumors: assessment of WT1 putative transcriptional targets using cDNA microarray expression profiling of genetically defined Wilms' tumors. Oncogene. 2003;22(24):3821-6. https://doi.org/10.1038/si.onc.1206597.
- 106. Rae FK, Martinez G, Gillinder KR, Smith A, Shooter G, Forrest AR, et al. Analysis of complementary expression profiles following WT1 induction versus repression reveals the cholesterol/fatty acid synthetic pathways as a possible major target of WT1. Oncogene. 2004;23(17):3067–79. https://doi.org/10.1038/sj.onc. 1207360.
- Graham K, Li W, Williams BR, Fraizer G. Vascular endothelial growth factor (VEGF) is suppressed in WT1-transfected LNCaP cells. Gene Expr. 2006;13(1):1–14. https://doi.org/10.3727/000000006783991953.
- Guo G, Morrison DJ, Licht JD, Quaggin SE. WT1 activates a glomerular-specific enhancer identified from the human nephrin gene. J Am Soc Nephrol. 2004;15(11):2851–6. https://doi.org/ 10.1097/01.asn.0000143474.91362.c4.
- Wagner N, Wagner KD, Theres H, Englert C, Schedl A, Scholz H. Coronary vessel development requires activation of the TrkB neurotrophin receptor by the Wilms' tumor transcription factor Wt1. Genes Dev. 2005;19(21):2631–42. https://doi.org/10.1101/gad.346405.
- Miyoshi Y, Ando A, Egawa C, et al. High expression of Wilms' tumor suppressor gene predicts poor prognosis in breast cancer patients. Clin Cancer Res. 2002;8(5):1167–71.
- 111. Zhao T, Zhou Y, Wu X, Peng Z, Gao P, Sun N, et al. Effect of transcription factor WT1 on triple-negative breast cancer metastasis through PFKFB4-mediated glycolysis. J Clin Oncol. 2024;42(16_suppl):e1314. https://doi.org/10.1200/JCO.2024.42. 16_suppl.e1314.
- 112. Zhang Y, Yan WT, Yang ZY, Li YL, Tan XN, Jiang J, et al. The role of WT1 in breast cancer: clinical implications, biological effects and molecular mechanism. Int J Biol Sci. 2020;16(8):1474–80. https://doi.org/10.7150/ijbs.39958.
- 113. Reizner N, Maor S, Sarfstein R, Abramovitch S, Welshons WV, Curran EM, et al. The WT1 Wilms' tumor suppressor gene product interacts with estrogen receptor-α and regulates IGF-I receptor gene transcription in breast cancer cells. J Mol Endocrinol. 2005;35(1):135–44. https://doi.org/10.1677/jme.1.01761.



- 114. Nasomyon T, Samphao S, Sangkhathat S, Mahattanobon S, Graidist P. Correlation of Wilms' tumor 1 isoforms with HER2 and ER-α and its oncogenic role in breast cancer. Anticancer Res. 2014;34(3):1333–42.
- 115. Müller M, Persson AB, Krueger K, Kirschner KM, Scholz H. The Wilms tumor protein WT1 stimulates transcription of the gene encoding insulin-like growth factor binding protein 5 (IGFBP5). Gene. 2017;619:21–9. https://doi.org/10.1016/j.gene.2017.03.037.
- 116. Bissanum R, Lirdprapamongkol K, Svasti J, Navakanitworakul R, Kanokwiroon K. The role of WT1 isoforms in vasculogenic mimicry and metastatic potential of human triple negative breast cancer cells. Biochem Biophys Res Commun. 2017;494(1–2):256–62. https://doi.org/10.1016/j.bbrc.2017.10.043.
- 117. Takeichi M, Nimura K, Mori M, Nakagami H, Kaneda Y. The transcription factors Tbx18 and Wt1 control the epicardial epithelial-mesenchymal transition through bi-directional regulation of Slug in murine primary epicardial cells. PLoS ONE. 2013;8(2):e57829. https://doi.org/10.1371/journal.pone.00578
- 118. Artibani M, Sims A, Slight J, Aitken S, Thornburn A, Muir M, et al. WT1 expression in breast cancer disrupts the epithelial/mesenchymal balance of tumour cells and correlates with the metabolic response to docetaxel. Sci Rep. 2017;7:45255. https://doi.org/10.1038/srep45255.
- Wu J, Yan H, Xiang C. Wilms' tumor gene 1 in hematological malignancies: friend or foe? Hematology. 2023;28(1):2254557. https://doi.org/10.1080/16078454.2023.2254557.
- Niktoreh N, Weber L, Walter C, Karimifard M, Hoffmeister LM, Breiter H, et al. Understanding WT1 alterations and expression profiles in hematological malignancies. Cancers (Basel). 2023;15(13):3491. https://doi.org/10.3390/cancers15133491.
- 121. Pan X, Mengge G, Wang K, Wang Y, Kong J, Sun Y, et al. Prognostic impact of WT1 mutation on AML of different risk groups based on 2022 European Leukemianet (ELN) risk classification. Blood. 2022;140(Suppl 1):3216–7. https://doi.org/10.1182/blood-2022-166323.
- 122. Koczkodaj D, Zmorzyoski S, Grygalewicz B, Pieokowska-Grela B, Styk W, Popek-Marciniec S, et al. WT1 gene mutations, rs16754 variant, and WT1 overexpression as prognostic factors in acute myeloid leukemia patients. J Clin Med. 2022;11(7):1873. https://doi.org/10.3390/jcm11071873.
- 123. Ghalandary M, Gao Y, Amend D, Kutkaite G, Vick B, Spiekermann K, et al. WT1 and DNMT3A play essential roles in the growth of certain patient AML cells in mice. Blood. 2023;141(8):955–60. https://doi.org/10.1182/blood.2022016411.
- Pan D, Zhao W, Jiang Q, et al. Wilms' tumor 1 expression combined with genetic mutations for prognostic assessment in MDS. Leuk Lymphoma. 2023;64(4):856–64. https://doi.org/10.1080/10428194.2023.2185086.
- 125. Xu C, Wu C, Xia Y, Zhong Z, Liu X, Xu J, et al. WT1 promotes cell proliferation in non-small cell lung cancer cell lines through up-regulating cyclin D1 and p-pRb in vitro and in vivo. PLoS ONE. 2013;8(8):e68837. https://doi.org/10.1371/journal.pone. 0068837.
- 126. Taube ET, Denkert C, Sehouli J, et al. Wilms tumor protein 1 (WT1)—not only a diagnostic but also a prognostic marker in high-grade serous ovarian carcinoma. Gynecol Oncol. 2016;140(3):494–502. https://doi.org/10.1016/j.ygyno.2015. 12.018.
- 127. Han Y, Song C, Zhang T, et al. Wilms' tumor 1 (WT1) promotes ovarian cancer progression by regulating E-cadherin and ERK1/2 signaling. Cell Cycle. 2020;19(20):2662–75. https://doi.org/10.1080/15384101.2020.1817666.
- Van Amerongen RA, Hagedoorn RS, Remst DFG, Assendelft DC, van der Steen DM, Wouters A, et al. WT1-specific TCRs

- directed against newly identified peptides install antitumor reactivity against acute myeloid leukemia and ovarian carcinoma. J Immunother Cancer. 2022;10(6):e004409. https://doi.org/10.1136/jitc-2021-004409.
- 129. Zeng Q, Liu T, Qin L, Wang C, Peng G, Li Z, et al. Screening of potential hub genes involved in kidney Wilms tumor via bioinformatics analysis and experimental validation. BMC Cancer. 2024;24(1):771. https://doi.org/10.1186/s12885-024-12541-x.
- Treger TD, et al. Stromal β-catenin activation impacts nephron progenitor differentiation in the developing kidney and may contribute to Wilms tumor. Development. 2020;147(13):dev190211. https://doi.org/10.1242/dev.190211.
- 131. Royer-Pokora B, Wruck W, Adjaye J, Beier M. Gene expression studies of WT1 mutant Wilms tumor cell lines in the framework of published kidney development data reveals their early kidney stem cell origin. PLoS ONE. 2023;18(1):e0270380. https://doi. org/10.1371/journal.pone.0270380.
- 132. Rialdi A, Duffy M, Scopton AP, et al. WNTinib is a multi-kinase inhibitor with specificity against β-catenin mutant hepatocellular carcinoma. Nat Cancer. 2023;4(8):1157–75. https://doi.org/10.1038/s43018-023-00609-0.
- 133. Gao H, et al. Emerging role of WT1 in cancer: a therapeutic target for solid tumors. Front Oncol. 2024;14:1032148. https://doi.org/10.3389/fonc.2024.1032148.
- 134. Angelico G, Santoro A, Straccia P, et al. Diagnostic and prognostic role of WT1 immunohistochemical expression in uterine carcinoma: a systematic review and meta-analysis across all endometrial carcinoma histotypes. Diagnostics. 2020;10(9):637. https://doi.org/10.3390/diagnostics10090637.
- 135. McEachron J, Baqir AW, Zhou N, et al. Evaluation of the incidence and clinical significance of WT-1 expression in uterine serous carcinoma. Gynecol Oncol Rep. 2021;39:100918. https://doi.org/10.1016/j.gore.2021.100918.
- Kanai T, Ito Z, Oji Y, et al. Prognostic significance of Wilms' tumor 1 expression in patients with pancreatic ductal adenocarcinoma. Oncol Lett. 2018;16(2):2682–92. https://doi.org/10.3892/ ol.2018.8961.
- 137. Qi XW, Zhang F, Wu H, et al. Wilms' tumor 1 (WT1) expression and prognosis in solid cancer patients: a systematic review and meta-analysis. Sci Rep. 2015;5:8924. https://doi.org/10.1038/srep08924.
- 138. Ota S, et al. Baseline immunity predicts prognosis of pancreatic cancer patients treated with WT1 and/or MUC1 peptide-loaded dendritic cell vaccination and a standard chemotherapy. Hum Vaccin Immunother. 2021;17(12):5563–72. https://doi.org/10.1080/21645515.2021.2003645.
- 139. Kim A, Park EY, Kim K, et al. Prognostic significance of WT1 expression in soft tissue sarcoma. World J Surg Oncol. 2014;12:214. https://doi.org/10.1186/1477-7819-12-214.
- Hashiba T, Izumoto S, Kagawa N, et al. Expression of WT1 protein and correlation with cellular proliferation in glial tumors.
 Neurol Med Chir (Tokyo). 2007;47(4):165–70. https://doi.org/10.2176/nmc.47.165.
- Manocha A, Jain S. WT1 in astrocytomas: comprehensive evaluation of immunohistochemical expression and its potential utility in different histological grades. Indian J Cancer. 2019;56(3):197–201. https://doi.org/10.4103/ijc.IJC_51_18.
- Mahzouni P, Meghdadi Z. WT1 protein expression in astrocytic tumors and its relationship with cellular proliferation index. Adv Biomed Res. 2013;2:33. https://doi.org/10.4103/2277-9175. 108772.
- 143. Chen R, Simpson DR, Zheng Y, Vaka D, Vicen ST, Sweet-Cordero A. Abstract PR08: Identifying effectors of WT1 that regulate KRAS-induced senescence and chemoresistance in non-small cell lung cancer. Mol Cancer Ther. 2013;12(5 Suppl):PR08. https://doi.org/10.1158/1535-7163.PMS-PR08.



- 144. Xu D, Chen X, Wu M, Bi J, Xue H, Chen H. Identification of cellular senescence-associated genes as new biomarkers for predicting the prognosis and immunotherapy response of non-small cell lung cancer and construction of a prognostic model. Heliyon. 2024;10(7):e28278. https://doi.org/10.1016/j.heliyon.2024. e28278.
- 145. Magrath JW, Sampath SS, Flinchum DA, Hartono AB, Goldberg IN, Boehling JR, et al. Comprehensive transcriptomic analysis of EWSR1::WT1 targets identifies CDK4/6 inhibitors as an effective therapy for desmoplastic small round cell tumors. Cancer Res. 2024;84(9):1426–42. https://doi.org/10.1158/0008-5472. CAN-23-3334.
- 146. Magrath JW, Espinosa-Cotton M, Flinchum DA, Sampath SS, Cheung NK, Lee SB. Desmoplastic small round cell tumor: from genomics to targets, potential paths to future therapeutics. Front Cell Dev Biol. 2024;12:1442488. https://doi.org/10.3389/fcell. 2024.1442488.
- 147. Renner M, Oleś M, Paramasivam N, Schneider A, Heilig CE, Modugno C, et al. 1936P Targeted therapy of desmoplastic small round cell tumor guided by multilayered molecular profiling. Ann Oncol. 2023;34(Suppl 2):S1032S1061. https://doi.org/10.1016/S0923-7534(23)01925-7.
- 148. Arai S, Tachibana T, Izumi A, et al. WT1-guided pre-emptive therapy after allogeneic hematopoietic stem cell transplantation in patients with acute myeloid leukemia. Int J Hematol. 2024;120(3):337–46. https://doi.org/10.1007/s12185-024-03795-z.
- 149. Tomlinson B, Lazarus HM. Enhancing acute myeloid leukemia therapy—monitoring response using residual disease testing as a guide to therapeutic decision-making. Expert Rev Hematol. 2017;10(6):563–74. https://doi.org/10.1080/17474086.2017. 1326811.
- 150. Schuurhuis GJ, Heuser M, Freeman S, Béné MC, Buccisano F, Cloos J, et al. Minimal/measurable residual disease in AML: a consensus document from the European LeukemiaNet MRD working party. Blood. 2018;131(12):1275–91. https://doi.org/10.1182/blood-2017-09-801498.
- 151. Sato H, Kobayashi T, Kameoka Y, et al. Prognostic impact of peripheral blood WT1 mRNA dynamics in patients with acute myeloid leukemia treated with venetoclax combination therapy. Int J Clin Oncol. 2024;29(4):481–92. https://doi.org/10.1007/ s10147-024-02480-9.
- 152. Li H, Xing C, Zhou B, Ye H, Feng J, Wu J, et al. A regulatory circuitry between miR-193a/miR-600 and WT1 enhances leuke-mogenesis in acute myeloid leukemia. Exp Hematol. 2018;61:59-68.e5. https://doi.org/10.1016/j.exphem.2018.02.001.
- 153. Nomdedéu JF, Esquirol A, Carricondo M, et al. Bone marrow WT1 levels in allogeneic hematopoietic stem cell transplantation for acute myelogenous leukemia and myelodysplasia: clinically relevant time points and 100 copies threshold value. Biol Blood Marrow Transplant. 2018;24(1):55–63. https://doi.org/10.1016/j.bbmt.2017.09.001.
- 154. Goel H, Rahul E, Gupta AK, et al. Molecular update on biology of Wilms Tumor 1 gene and its applications in acute myeloid leukemia. Am J Blood Res. 2020;10(5):151–60.
- 155. Messina S, Tresoldi C, Stanghellini MTL, Crotta A, et al. WT1 transcripts is a powerful leukemia marker to predict early relapse after allogeneic haematopoietic stem cell transplantation. Blood. 2009;114(22):4488. https://doi.org/10.1182/blood.V114.22.4488.4488.
- 156. Hirahata T, Ul Quraish R, Quraish AU, Ul Quraish S, Naz M, Razzaq MA. Liquid biopsy: a distinctive approach to the diagnosis and prognosis of cancer. Cancer Inform. 2022;21:11769351221076062. https://doi.org/10.1177/11769351221076062.

- 157. Cilloni D, Renneville A, Hermitte F, et al. Real-time quantitative polymerase chain reaction detection of minimal residual disease by standardized WT1 assay to enhance risk stratification in acute myeloid leukemia: a European LeukemiaNet study. J Clin Oncol. 2009:27(31):5195–201.
- 158. Lambert J, Thomas X, Marceau-Renaut A, Micol JB, Renneville A, et al. Early detection of WT1 measurable residual disease identifies high-risk patients, independent of transplantation in AML. Blood Adv. 2021;5(23):5258–68. https://doi.org/10.1182/ bloodadvances.2021004322.
- Lazzarotto D, Candoni A. The role of Wilms' tumor gene (WT1) expression as a marker of minimal residual disease in acute myeloid leukemia. J Clin Med. 2022;11(12):3306. https://doi.org/10.3390/jcm11123306.
- Ghannam J, Dillon LW, Hourigan CS. Next-generation sequencing for measurable residual disease detection in acute myeloid leukaemia. Br J Haematol. 2020;188(1):77–85. https://doi.org/10.1111/bjh.16362.
- 161. Namdaroğlu S, Başcı S, Aslan Candır B, Yaman S, Yiğenoğlu TN, Bahsi T, et al. Role of WT1 in measurable residual disease follow-up in the post allogeneic stem cell transplant setting. J Clin Med. 2024;13(17):5145. https://doi.org/10.3390/jcm13 175145.
- 162. Donia HM, Elsweify NM, Farahat NM, Nadwan EA. Wilms tumor 1 gene expression in acute myeloid leukemia: prognostic significance and usefulness in minimal residual disease monitoring—a case—control study. Egypt J Med Hum Genet. 2022;23:109. https://doi.org/10.1186/s43042-022-00291-z.
- 163. Ugale PR, Achrekar AR, Biswas S, Joshi S, Terse V, et al. Prognostic significance of Wilms' tumor 1 gene mutations in acute myeloid leukemia: risk stratification and survival outcomes. Blood. 2024;144(1):1550. https://doi.org/10.1182/ blood-2024-199066.
- 164. Coosemans A, Van Calster B, Verbist G, et al. Wilms tumor gene 1 (WT1) is a prognostic marker in high-grade uterine sarcoma. Int J Gynecol Cancer. 2011;21(2):302–8. https://doi. org/10.1097/IGC.0b013e318207cab5.
- 165. Guo W, Zhang H, Wang M, Zheng Y, Cao Y, et al. WT1 gene mutations impact post-transplant relapse in myelo-dysplastic syndrome with excess blasts 2 patients. Ann Hematol. 2024;103(8):2827-36. https://doi.org/10.1007/s00277-024-05870-1.
- 166. Ahmed S, Elsherif M, Yassin D, et al. Integration of measurable residual disease by WT1 gene expression and flow cytometry identifies pediatric patients with high risk of relapse in acute myeloid leukemia. Front Oncol. 2024;14:1340909. https://doi.org/10.3389/fonc.2024.1340909.
- 167. Oka Y, Tsuboi A, Murakami M, Hirai M, Tominaga N, Nakajima H, et al. Wilms tumor gene peptide-based immunotherapy for patients with overt leukemia from myelodysplastic syndrome (MDS) or MDS with myelofibrosis. Int J Hematol. 2003;78(1):56–61. https://doi.org/10.1007/BF02983241.
- Sugiyama H. SY7–6-WT1 peptide-based cancer immunotherapy. Ann Oncol. 2012;23(Suppl 11):xi74. https://doi.org/10. 1016/S0923-7534(20)32149-9.
- Karam S, Gebreil A, Alksas A, et al. Insights into personalized care strategies for Wilms tumor: a narrative literature review. Biomedicines. 2024;12(7):1455. https://doi.org/10.3390/biomedicines12071455.
- Ogasawara M. Wilms' tumor 1-targeting cancer vaccine: recent advancements and future perspectives. Hum Vaccin Immunother. 2024;20(1):2296735. https://doi.org/10.1080/21645 515.2023.2296735.
- 171. Naoe T, Saito A, Hosono N, et al. Immunoreactivity to WT1 peptide vaccine is associated with prognosis in elderly patients with acute myeloid leukemia: follow-up study of randomized



- phase II trial of OCV-501, an HLA class II-binding WT1 polypeptide. Cancer Immunol Immunother. 2023;72(8):2865–71. https://doi.org/10.1007/s00262-023-03432-4.
- 172. Hashii Y. WT1-targeting immunotherapy in children with refractory cancer. Jpn J Pediatr Hematol Oncol. 2021;58(3):208-12. https://doi.org/10.11412/jspho.58.208.
- 173. Tsuboi A, Hashimoto N, Fujiki F, et al. A phase I clinical study of a cocktail vaccine of Wilms' tumor 1 (WT1) HLA class I and II peptides for recurrent malignant glioma. Cancer Immunol Immunother. 2019;68(2):331–40. https://doi.org/10.1007/ s00262-018-2274-1.
- 174. Gugulothu KN, Anvesh Sai P, Suraparaju S, Karuturi SP, Pendli G, Kamma RB, et al. WT1 cancer vaccine in advanced pancreatic cancer: a systematic review. Cureus. 2024;16(3):e56934. https://doi.org/10.7759/cureus.56934.
- 175. Kreutmair S, Pfeifer D, Waterhouse M, et al. First-in-human study of WT1 recombinant protein vaccination in elderly patients with AML in remission: a single-center experience. Cancer Immunol Immunother. 2022;71(12):2913–28. https:// doi.org/10.1007/s00262-022-03202-8.
- 176. Suwabe T, Shibasaki Y, Tamura S, Katagiri T, Fuse K, Ida-Kurasaki T, et al. Decade-long WT1-specific CTLs induced by WT1 peptide vaccination. Int J Hematol. 2024;119(4):399– 406. https://doi.org/10.1007/s12185-024-03723-1.
- 177. Van Driessche A, Berneman ZN, Van Tendeloo VF. Active specific immunotherapy targeting the Wilms' tumor protein 1 (WT1) for patients with hematological malignancies and solid tumors: lessons from early clinical trials. Oncologist. 2012;17(2):250–9. https://doi.org/10.1634/theoncologist.2011-0240.
- Bawek S, Gurusinghe S, Burwinkel M, Przespolewski A. Updates in novel immunotherapeutic strategies for relapsed/refractory AML. Front Oncol. 2024;14:1374963. https://doi.org/10.3389/ fonc.2024.1374963.
- 179. Versteven M, Damoiseaux D, Campillo-Davo D, Van Acker H. Abstract B137: Preclinical evaluation of a Wilms' tumor protein 1-targeted interleukin-15 dendritic cell vaccine: T-cell activity and batch production. Cancer Immunol Res. 2019;7(2 Suppl):B137. https://doi.org/10.1158/2326-6074.CRICIMTEAT IAACR18-B137.
- 180. Katsuda M, Miyazawa M, Ojima T, Hagiwara S, Takahashi R, Yamashita K, et al. A double-blind randomized comparative clinical trial to evaluate the safety and efficacy of dendritic cell vaccine loaded with WT1 peptides (TLP0001) in combination with S-1 in patients with advanced pancreatic cancer refractory to standard chemotherapy. Trials. 2019;20(1):242. https://doi.org/10.1186/s13063-019-3332-5.
- 181. Manning-Geist BL, Gnjatic S, Aghajanian C, Levine DA, Iasonos A, Sabbatini PJ, et al. Phase I study of a multivalent WT1 peptide vaccine (Galinpepimut-S) in combination with nivolumab in patients with WT1-expressing ovarian cancer in second or third remission. Cancers (Basel). 2023;15(5):1458. https://doi.org/10.3390/cancers15051458.
- 182. Takahashi M. Long-term molecular remission with an increase of WT1-specific CTLs after cessation of WT1 peptide vaccination in a patient with CML. Ann Oncol. 2015;26(Suppl 2):ii23. https://doi.org/10.1093/annonc/mdv093.4.
- 183. Ohno S, Okuyama R, Aruga A, Sugiyama H, Yamamoto M. Phase I trial of Wilms' tumor 1 (WT1) peptide vaccine with GM-CSF or CpG in patients with solid malignancy. Anticancer Res. 2012;32(6):2263–9.
- 184. Kuball J, de Boer K, Wagner E, Wattad M, Antunes E, Weeratna RD, et al. Pitfalls of vaccinations with WT1-, Proteinase3- and MUC1-derived peptides in combination with Montanide ISA51 and CpG7909. Cancer Immunol Immunother. 2011;60(2):161–71. https://doi.org/10.1007/s00262-010-0929-7.

- 185. Nakagawa N, Hashii Y, Kayama H, Okumura R, Nakajima H, Minagawa H, et al. An oral WT1 protein vaccine composed of WT1-anchored, genetically engineered Bifidobacterium longum allows for intestinal immunity in mice with acute myeloid leukemia. Cancer Immunol Immunother. 2023;72(1):39–53. https://doi.org/10.1007/s00262-022-03214-4.
- 186. Minagawa H, Hashii Y, Nakajima H, Fujiki F, Morimoto S, Nakata J, et al. Enhanced antitumor activity of a novel, oral, helper epitope-containing WT1 protein vaccine in a model of murine leukemia. BMC Cancer. 2023;23(1):167. https://doi.org/10.1186/s12885-023-10547-5.
- 187. Cho SY, Jeong SM, Jeon YJ, Yang S, Hwang JE, Lim SH, et al. WT1 pulsed human CD141+ dendritic cell vaccine has high potential in solid tumor-targeted immunotherapy. Int J Mol Sci. 2023;24(2):1501. https://doi.org/10.3390/ijms24021501.
- 188. Nishida S, Ishikawa T, Egawa S, Shimizu S, Takano S, Aoki T, et al. Combination gemcitabine and WT1 peptide vaccination improves progression-free survival in advanced pancreatic ductal adenocarcinoma: a phase II randomized study. Cancer Immunol Res. 2018;6(3):320–31. https://doi.org/10.1158/2326-6066. CIR-17-0386.
- 189. Higgins M, Curigliano G, Dieras V, Letarte N, Sahmoud T, Humbert M, et al. Safety and immunogenicity of neoadjuvant treatment using WT1-immunotherapeutic in combination with standard therapy in patients with WT1positive stage II/III breast cancer: a randomized phase I study. Breast Cancer Res Treat. 2017;162(3):479–88. https://doi.org/10.1007/s10549-017-4130-y.
- 190. Walters JN, Ferraro B, Duperret EK, Clark A, Espinosa DA, Krasnow NA, et al. A novel DNA vaccine platform enhances neo-antigen-like T cell responses against WT1 to break tolerance and induce anti-tumor immunity. Mol Ther. 2017;25(4):976–88. https://doi.org/10.1016/j.ymthe.2017.01.022.
- 191. Zhang X, Goedegebuure SP, Chen MY, Abbott KL, Hartman ZC, Lin JR, et al. Neoantigen DNA vaccines are safe, feasible, and induce neoantigen-specific immune responses in triple-negative breast cancer patients. Genome Med. 2024;16(1):131. https://doi. org/10.1186/s13073-024-01388-3.
- 192. Gao S, Yang L, Du J, Jiang W, Fang L. Abstract 6705: a novel tri-specific T cell engager targeting the intracellular oncoprotein WT1. Cancer Res. 2024;84(6 Suppl):6705. https://doi.org/10. 1158/1538-7445.AM2024-6705.
- 193. Biocytogen Pharmaceuticals (Beijing) Co., Ltd, CtM Biotech (Shanghai) Co., Ltd. Biocytogen and CtM Bio achieve milestone progress in co-developing a TCR-mimic antibody-based multispecific T cell engager targeting WT1/HLA-A02 [Internet]. Beijing and Shanghai (China): Biocytogen; 2023 Dec 27 [cited 2025 Jun 27]. Available from: https://www.biocytogen.com/news/ctm-wt1-milestone
- 194. Dubrovsky L, Pankov D, Brea EJ, Malavasi F, Lenz HJ, Junghans RP, et al. A TCR-mimic antibody to WT1 bypasses tyrosine kinase inhibitor resistance in human BCR-ABL+ leukemias. Blood. 2014;123(21):3296–304. https://doi.org/10.1182/blood-2014-01-549022.
- 195. Shen Y, Li YM, Zhou JJ, Lu H, Wang F, Zhang Y, et al. The antitumor activity of TCR-mimic antibody-drug conjugates (TCRm-ADCs) targeting the intracellular Wilms tumor 1 (WT1) oncoprotein. Int J Mol Sci. 2019;20(16):3912. https://doi.org/10.3390/ijms20163912.
- 196. Augsberger C, Hänel G, Xu W, Gans S, Bräutigam J, Vogel W, et al. Targeting intracellular WT1 in AML with a novel RMF-peptide-MHC-specific T-cell bispecific antibody. Blood. 2021 Dec 23;138(25):2655–69. Correction in: Blood. 2022 Mar 31; 139(13):2086–7. https://doi.org/10.1182/blood.2022015672.



- 197. Mun SS, Meyerberg J, Peraro L, Benmebarek MR, Touzot M, Sebbagh R, et al. Dual targeting ovarian cancer by MUC16 CAR T cells secreting a bispecific T cell engager antibody for an intracellular tumor antigen WT1. Cancer Immunol Immunother. 2023;72(11):3773–86. https://doi.org/10.1007/s00262-023-03529-w.
- Zhang Y, Li Y. T cell receptor-engineered T cells for leukemia immunotherapy. Cancer Cell Int. 2019;3(19):2. https://doi.org/ 10.1186/s12935-018-0720-y.
- 199. Gielis S, Flumens D, van der Heijden S, van Acker H, Peeters M, Willems E, et al. Analysis of Wilms' tumor protein 1 specific TCR repertoire in AML patients uncovers higher diversity in patients in remission than in relapsed. Ann Hematol. 2025;104(1):317–33. https://doi.org/10.1007/ s00277-024-05919-1.
- Tawara I, Kageyama S, Miyahara Y, Fujiwara H, Nishida T, Chiba S, et al. Safety and persistence of WT1-specific T-cell receptor gene-transduced lymphocytes in patients with AML and MDS. Blood. 2017;130(18):1985–94. https://doi.org/10.1182/ blood-2017-06-791202.
- Kawamoto H, Masuda K, Nagano S. Regeneration of antigenspecific T cells by using induced pluripotent stem cell (iPSC) technology. Int Immunol. 2021;33(12):827–33. https://doi.org/ 10.1093/intimm/dxab091.
- Ruggiero E, Carnevale E, Prodeus A, Magnani ZI, Camisa B, Merelli I, et al. CRISPR-based gene disruption and integration of high-avidity, WT1-specific T cell receptors improve antitumor T cell function. Sci Transl Med. 2022;14:eabg8027. https://doi. org/10.1126/scitranslmed.abg8027.
- Naghizadeh S, Mansoori B, Mohammadi A, Sakhinia E, Baradaran B. Gene silencing strategies in cancer therapy: an update for drug resistance. Curr Med Chem. 2019;26(34):6282–300. https://doi.org/10.2174/0929867325666180403141554.
- 204. Graziano AC, Cardile V, Avola R, Tomasello L, Battiato C, Giunta E, et al. Wilms' tumor gene 1 silencing inhibits proliferation of human osteosarcoma MG-63 cell line by cell cycle arrest and apoptosis activation. Oncotarget. 2017;8(8):13917–31. https://doi.org/10.18632/oncotarget.14715.
- 205. Elmaagacli AH, Koldehoff M, Peceny R, Trenschel R, Beelen DW, Hallek M, et al. WT1 and BCR-ABL specific small interfering RNA have additive effects in the induction of apoptosis in leukemic cells. Haematologica. 2005;90(3):326–34.
- Glienke W, Maute L, Koehl U, Esser R, Milz E, Bergmann L. Effective treatment of leukemic cell lines with WT1 siRNA. Leukemia. 2007;21(10):2164–70. https://doi.org/10.1038/sj. leu.2404878.
- 207. Maslak PG, Dao T, Bernal Y, Smith JS, O'Rourke K, Cooper LJ, et al. Phase 2 trial of a multivalent WT1 peptide vaccine (galinpepimut-S) in acute myeloid leukemia. Blood Adv. 2018;2(3):224–34. https://doi.org/10.1182/bloodadvances. 2017014175.
- 208. Benteyn D, Anguille S, Van Lint S, Heirman C, Van Overmeire E, Berghen C, et al. Design of an optimized Wilms' tumor 1 (WT1) mRNA construct for enhanced WT1 expression and improved immunogenicity in vitro and in vivo. Mol Ther Nucleic Acids. 2013;2(11):e134. https://doi.org/10.1038/mtna. 2013.54.
- 209. Van Genechten T, De Laere M, Van den Bossche J, Goovaerts T, Peeters M, Van Camp B, et al. Adjuvant Wilms' tumour 1-specific dendritic cell immunotherapy complementing conventional therapy for paediatric patients with high-grade glioma and diffuse intrinsic pontine glioma: protocol of a monocentric phase I/II clinical trial in Belgium. BMJ Open. 2024;14(3):e077613. https://doi.org/10.1136/bmjopen-2023-077613.

- Zhao Q, Ahmed M, Tassev DV, Savoldo B, Majumder P, Cruz CRY, et al. Affinity maturation of T-cell receptor-like antibodies for Wilms tumour 1 peptide greatly enhances therapeutic potential. Leukemia. 2015;29(11):2238–47. https://doi.org/10.1038/leu.2015.125.
- 211. Zamora-Avila DE, Franco-Molina MA, Trejo-Avila LM, Rodríguez-Padilla C, Reséndez-Pérez D, Zapata-Benavides P. RNAi silencing of the WT1 gene inhibits cell proliferation and induces apoptosis in the B16F10 murine melanoma cell line. Melanoma Res. 2007;17(6):341–8. https://doi.org/10.1097/CMR.0b013e3282efd3ae.
- 212. Tatsumi N, Oji Y, Tsuji N, Ohno T, Sugiyama H, Komatsu T, et al. Wilms' tumor gene WT1-shRNA as a potent apoptosis-inducing agent for solid tumors. Int J Oncol. 2008;32(3):701-11.
- 213. Loeb DM, Sukumar S. The role of WT1 in oncogenesis: tumor suppressor or oncogene? Int J Hematol. 2002;76(2):117–26. https://doi.org/10.1007/BF02982573.
- 214. Wang X, Zhao J, Zhang Y, He Y, Li G, Chen H, et al. Molecular mechanism of Wilms' tumor (Wt1) +/-KTS variants promoting proliferation and migration of ovarian epithelial cells by bioinformatics analysis. J Ovarian Res. 2023;16(1):46. https://doi.org/10.1186/s13048-023-01124-2.
- 215. Calabrese C, Pica GM, Bracco E, Serra A, D'Amico G, Battaglia M, et al. Detection of an unbalanced ratio between WT1 isoforms in acute myeloid leukemia and its correlation with molecular abnormalities. Blood. 2012;120(21):2500. https://doi.org/10.1182/blood.V120.21.2500.250.
- Hashii Y, Sato E, Ohta H, Oka Y, Sugiyama H, Ozono K. WT1 peptide immunotherapy for cancer in children and young adults. Pediatr Blood Cancer. 2010;55(2):352–5. https://doi. org/10.1002/pbc.22522.
- 217. Kiguchi T, Yamaguchi M, Takezako N, Yamamoto K, Tsujita T, Tanaka J, et al. Efficacy and safety of Wilms' tumor 1 helper peptide OCV-501 in elderly patients with acute myeloid leukemia: a multicenter, randomized, double-blind, placebo-controlled phase 2 trial. Cancer Immunol Immunother. 2022;71(6):1419–30. https://doi.org/10.1007/s00262-02103
- 218. Matsuda T, Takeuchi H, Sakurai T, Komori T, Yoshida K, Mori T, et al. Pilot study of WT1 peptide-pulsed dendritic cell vaccination with docetaxel in esophageal cancer. Oncol Lett. 2018;16(1):1348–56. https://doi.org/10.3892/ol.2018.8734.
- Mayanagi S, Kitago M, Sakurai T, Komori T, Mori T, Saisho S, et al. Phase I pilot study of Wilms tumor gene 1 peptide-pulsed dendritic cell vaccination combined with gemcitabine in pancreatic cancer. Cancer Sci. 2015;106(4):397–406. https://doi.org/10.1111/cas.12621.
- Kitagawa K, Gonoi R, Tatsumi M, Matsushita H, Mori T, Saisho S, et al. Preclinical development of a WT1 oral cancer vaccine using a bacterial vector to treat castration-resistant prostate cancer. Mol Cancer Ther. 2019;18(5):980–90. https://doi.org/10.1158/1535-7163.MCT-18-1105.
- 222. Hashii Y, Oka Y, Kagawa N, Komori T, Hashimoto H, Hashimoto Y, et al. Encouraging clinical evolution of a pediatric patient with relapsed diffuse midline glioma who underwent WT1-targeting immunotherapy: a case report and literature review. Front Oncol. 2020;24(10):1188. https://doi.org/10.3389/fonc.2020.01188.
- 223. Hosen N, Maeda T, Hashii Y, Kawaguchi R, Komori T, Saisho S, et al. Wilms tumor 1 peptide vaccination after hematopoietic



- stem cell transplant in leukemia patients. Stem Cell Investig. 2016;3:90. https://doi.org/10.21037/sci.2016.11.08.
- 224. Van Lint S, Goyvaerts C, Maenhout S, Heirman C, Van der Bruggen P, Thielemans K, et al. Preclinical evaluation of Tri-Mix and antigen mRNA-based antitumor therapy. Cancer Res. 2012;72(7):1661–71. https://doi.org/10.1158/00085472. CAN-11-2957.
- 225. De Keersmaecker B, Claerhout S, Carrasco J, Anguille S, Van Lint S, Van den Oord J, et al. TriMix and tumor antigen mRNA electroporated dendritic cell vaccination plus ipilimumab: link between T-cell activation and clinical responses in advanced melanoma. J Immunother Cancer. 2020;8(1):e000329. https://doi.org/10.1136/jitc-2019-000329.
- 226. Chen M, Fang X, Du R, Zhang Y, Li Q, Zhao B, et al. A nucleus-targeting WT1 antagonistic peptide encapsulated in polymeric nanomicelles combats refractory chronic myeloid leukemia. Pharmaceutics. 2023;15(9):2305. https://doi.org/10.3390/pharmaceutics15092305.
- Xie YJ, Liu WQ, Li D, Hou JC, Coghi PS, Fan XX. Overcoming suppressive tumor microenvironment by vaccines in solid tumor. Vaccines (Basel). 2023;11(2):394. https://doi.org/10.3390/vaccines11020394.
- Suzuki M, Hishida T, Asakura K, Asamura H. WT1 peptide-based immunotherapy for refractory thymic epithelial malignancies. Mediastinum. 2019;15(3):12. https://doi.org/10.21037/med. 2019.04.01
- 229. Morimoto S, Tanaka Y, Nakata J, Takayama T, Saisho S, Komori T, et al. Spontaneous high clonal expansion of Wilms' tumor gene 1-specific cytotoxic T-lymphocytes in patients with Wilms' tumor gene 1-expressing solid tumor. Cancer Immunol Immunother. 2024;74(1):15–23. https://doi.org/10.1007/s00262-024-03862-8.
- 230. Ito M, Koido S, Iwamoto T, Komori T, Saisho S, Kawashima H, et al. Enhancing the immunogenicity of Wilms tumor 1 epitope in mesothelioma cells with immunoproteasome inhibitors. PLoS ONE. 2024;19(8):e0308330. https://doi.org/10.1371/journal.pone.0308330.

- 231. Lahman MC, Schmitt TM, Paulson KG, Segal NH, Lim M, Spitz FR, et al. Targeting an alternate Wilms' tumor antigen 1 peptide bypasses immunoproteasome dependency. Sci Transl Med. 2022;14(631):eabg8070. https://doi.org/10.1126/scitranslmed.abg8070.
- 232. Deng DX, Wen JJ, Cheng YF, Yang WZ, Li YM, Liu Y, et al. Wilms' tumor gene 1 is an independent prognostic factor for pediatric acute myeloid leukemia following allogeneic hematopoietic stem cell transplantation. BMC Cancer. 2021;21(1):292. https://doi.org/10.1186/s12885-021-08022-0.
- 233. Boublikova L, Kalinova M, Ryan J, Trka J, Semeradova L, Chroma J, et al. Wilms' tumor gene 1 (WT1) expression in childhood acute lymphoblastic leukemia: a wide range of WT1 expression levels, its impact on prognosis and minimal residual disease monitoring. Leukemia. 2006;20(2):254–63. https://doi.org/10.1038/sj.leu.2404047.
- 234. Xue Y, Wang Y, Zhang LP, Chen L, Li J, Zhao J, et al. Prognostic significance of Wilms' tumor gene 1 expression in children with B-cell precursor acute lymphoblastic leukemia. Front Oncol. 2024;15(13):1297870. https://doi.org/10.3389/fonc.2023.1297870.
- 235. Osada T, Woo CY, McKinney M, Younes A, Takita H, Yoshimoto M, et al. Induction of Wilms' tumor protein (WT1)specific antitumor immunity using a truncated WT1-expressing adenovirus vaccine. Clin Cancer Res. 2009;15(8):2789–96. https://doi.org/10.1158/1078-0432.CCR-08-2589.
- Morishita Y, Takenouchi K, Sakashita S, Matsuura K, Hayashi R, Nakatsura T. Immunohistochemical analysis of common cancer antigens in head and neck squamous cell carcinoma. Anticancer Res. 2022;42(12):5751–61. https://doi.org/10.21873/anticanres. 16082.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

